Abstract

Recombinant, live, attenuated viruses of the Pneumoviridae family are disclosed that include a baculovirus GP64 envelope glycoprotein or variant or fragment thereof and a respiratory syncytial virus (RSV) F protein variant or fragment thereof. Also disclosed are polynucleotides encoding the virus as well as pharmaceutical compositions and vaccines containing the virus. In addition, methods of producing and using each of the above compositions are also disclosed.

22 Claims, 12 Drawing Sheets

Specification includes a Sequence Listing.
wildtype F

preF<sub>ΔCT</sub>
preF<sub>SEC</sub>
preF<sub>SEC/tag</sub>

TMD

574

S155C S190F V207L C062S

(Δ554-574)
(Δ514-574)
(Δ514-574/tag)

FIG. 1
FIG. 2
FIG. 3
D25: pre-fusion F specific (site 0)
14402: pre-fusion F specific (site V)
Motavizumab: recognizes pre- and post-fusion F
15576: post-fusion F specific
Anti-myc Ab: NGFR-myc is transfection control

FIG. 4
Relative surface expression (OD$_{490}$)

- RSV6-preF$^{ACT}$
- RSV rec WT
- RSV6-preF$^{SEC}$
- Mock

- D25: pre-fusion F specific (site 0)
- 14402: pre-fusion F specific (site V)
- AM14: recognizes only prefusion F in trimeric form
- 15576: post-fusion F specific
- Anti-G Ab (L9)
- Anti-N Ab (for normalization based on virus genome replication)

**FIG. 5**
FIG. 7

- **RSV6-preF<sup>ΔCT</sup>: 1 million PFU/vaccination**
- **RSV6-preF<sup>ΔCT</sup>: 0.5 million PFU/vaccination**
- **Rec WT RSV: 1 million PFU/vaccination**
- **Rec WT RSV: 0.5 million PFU/vaccination**
- **Serum from mock-vaccinated mice**
FIG. 9
FIG. 10

- Pooled sera from RSV6-preF<sup>ACT</sup> infected mice
- Pooled sera from RSV1-preF<sup>ACT</sup>-NS1<sup>low</sup> infected mice
- Pooled sera from Mock-infected mice
- Anti-G Antibody (L9)
Dilution that gives 50% neutralization (F-specific) (500 PFU)

Rec WT: 1/42
RSV1-preF<sup>ACT</sup>: 1/90
RSV1-preF<sup>ACT</sup>-NS1low: 1/80

FIG. 11
Dilution that gives 50% neutralization (F-specific) (250 PFU)

Rec WT: 1/55
RSV6-preF<sup>ACT</sup>: 1/540

FIG. 12
RSV VACCINES AND METHODS OF PRODUCTION AND USE THEREOF

CROSS REFERENCE TO RELATED APPLICATIONS/INCORPORATION BY REFERENCE STATEMENT

This application is a continuation of U.S. Ser. No. 17/015, 610, filed Sep. 9, 2020, which is a continuation of U.S. Ser. No. 16/257,738, filed Jan. 25, 2019, now U.S. Pat. No. 10,799,576, issued Oct. 13, 2020, which claims benefit under 35 USC § 119(e) of provisional application U.S. Ser. No. 62/621,685, filed Jan. 25, 2018. The entire contents of each of the above-referenced patent(s)/application(s) are expressly incorporated herein by reference.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This invention was made with Government support under Grant No. 1R21AI128520-01A1 awarded by the National Institutes of Health. The Government has certain rights in the invention.

BACKGROUND

Respiratory Syncytial Virus (RSV) is the single largest viral cause of pediatric bronchiolitis and pneumonia, with a high worldwide mortality. In spite of many years of clinical trials and scientific progress, a safe and effective vaccine against RSV has still not been found. In the 1960s, a formalin-inactivated RSV vaccine (FI-RSV) induced an imbalance in the immune response which led to enhanced pathology after exposure to wild type RSV (known as vaccine-enhanced disease (VED)). Ever since this encounter with VED, it has been enormously challenging to impart both sufficient safety and efficacy in a single vaccine. Furthermore, there are age-specific challenges, and it is generally believed that different vaccine platforms will be needed for different populations and/or age groups to lessen the RSV-associated disease burden. For RSV-naive children, live-attenuated vaccines are an important focus, because inactivated and subunit vaccines are poor at inducing cell-mediated immunity, and this is known to contribute to VED. Moreover, live vaccines typically can also induce broad systemic and local immunity. Thus, for RSV-naive individuals, a live vaccine approach is an attractive option, provided the vaccine itself is sufficiently safe and cannot revert to a more aggressive phenotype.

RSV contains a negative-sense, single-stranded RNA genome that expresses eleven known proteins from ten genes (FIG. 2). Of these, the attachment (A) and fusion (F) proteins have been characterized as transmembrane (surface) glycoproteins and contain the major antigenic epitopes of human respiratory syncytial virus; as such, the G and F proteins appear to be critical for induction of neutralizing anti-RSV antibodies. In contrast to G, F is essential for virus infectivity.

U.S. Pat. No. 7,588,770 to Oomens et al., the entire contents of which are hereby expressly incorporated herein by reference, describes genetically modified RSVs generated by replacing genes encoding proteins such as F and G with genes encoding heterologous envelope proteins, e.g., a baculovirus GP64 envelope glycoprotein. Such genetically modified RSVs exhibit improved temperature stability and in some cases are infectious but incapable of cell-to-cell transmission. Thus, these attenuated viruses are safe for use in vaccines. However, a disadvantage of this technology is that removal of the F and G proteins from the virus greatly reduces antigenicity, thereby decreasing the ability of the virus to elicit a robust, protective immune response.

It has recently been recognized that the viral fusion (F) protein is unstable and readily shifts to the post-fusion conformation during purification or vaccine preparation. As a result, a large proportion of vaccine-induced antibodies (Abs) target the post-fusion form, which is functionally obsolete. To avoid induction of anti-post-fusion F Abs, McLellan et al. were able to genetically stabilize the pre-fusion form (referred to as Pref), thereby greatly increasing neutralizing capacity of anti-F Abs when given as a protein vaccine (see, for example, US Patent Application Nos. US 2015/00306622 (published Jan. 29, 2015 to Marshall et al.); US 2016/0031972 (published Feb. 4, 2016 to Zheng et al.); and US 2016/0046675 (published Feb. 18, 2016 to Kwong et al.); the entire contents of which are hereby expressly incorporated herein by reference).

However, subunit vaccines are deemed unsafe for the RSV-naive target population. In addition, stabilization of Pref renders it non-functional, and a virus solely expressing Pref is not viable.

Therefore, there is a need in the art for new and improved RSV vaccines that overcome the disadvantages and defects of the prior art. It is to such new and improved vaccines, as well as methods of production and use thereof, that the present disclosure is directed.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 illustrates three versions of pre-fusion stabilized F protein variants that were generated for use in accordance with the present disclosure. These pre-fusion stabilized F protein variants are based on the previously described prefusion protein variant DS-Cav-1 (see, for example, US 2015/00306622, US 2016/0031972, and US 2016/0046675, incorporated supra; and McLellan et al. (Science (2013) 342:592-598); the entire contents of which are expressly incorporated herein by reference). pref<sup>DSCT</sup> is a membrane-anchored version that is expressed and anchored at the surface of infected cells. RSV-pref<sup>DSCT</sup> is a secreted version that is secreted to the extracellular environment on infected cells. RSV-pref<sup>DSCTag</sup> is similar to RSV-pref<sup>DSCT</sup> but contains an epitope tag for easy identification and detection. The DS-Cav-1 mutations are shown by vertical lines. A wildtype F ORF is shown for comparison (574 amino acids). TMD=transmembrane domain. Tag=epitope tag.

FIG. 2 graphically depicts the engineering of RSV viruses with pre-fusion stabilized F variants at the 8th or 6th genome position. Panel A, the wildtype RSV genome; Panel B, RSV genomes with variants of pre-fusion stabilized F at the 8th genome position; and Panel C, RSV genomes with variants of pre-fusion stabilized F at the 6th genome position. The 6th genome position is more highly expressed than the 8th genome position, to enhance the level of pre-fusion F. All viruses have a GFP marker gene for tracking and assay purposes. GFP is not required and can be removed if necessary. All viruses also lack expression of the secreted G protein (indicated as Gimen), which is a known virulence factor.

FIG. 3 graphically depicts that removal of the cytoplasmic tail (CT) strongly improved cell-surface expression of pre-fusion F. HEp2 cells were transfected with the indicated F expressing plasmids. The F open reading frames were codon-optimized, as native F sequences express poorly in transfected cells. To each well, a plasmid expressing NGFR-
myc was added as a transfection control (NGFR-myc is expressed at the cell surface). At 48 hour post-transfection, transfected cells were incubated with various F antibodies or myc antibody as a control, and relative F surface levels were detected using standard ELISA.

FIG. 4 graphically depicts verification of the engineered viruses generated as in FIG. 2 as successfully expressing pre-fusion stabilized F protein variants. Cells infected by virus RS8V-preF\(^{\text{ACCT}}\) were incubated with anti-F and anti-G antibodies at 26 hours post-infection, or mock-infected as a negative control, and subjected to ELISA. Three antibodies (provided by J S McEwan) were used to detect F. The first, Motavizumab (mota), detects both the pre-fusion and post-fusion conformation of F; the second and third antibodies (D25 and 14402) are known to detect a different epitope specific only for the pre-fusion conformation of F (site O and site V.G; 465 autoantibody in post-fusion levels. Abundant levels of pre-fusion F were expressed at the surface of vaccine-virus infected cells. All F Abs were applied at 0.1 µg/ml. Error bars are standard deviation of the mean from triplicate samples. Viruses RS8V-preF\(^{\text{ACCT}}\) and RS8V-preF\(^{\text{ACCT}}\) have been similarly examined and also express pre-fusion F.

FIG. 5 graphically depicts that vaccine candidate RS6-preF\(^{\text{ACCT}}\) induced high surface levels of pre-fusion F and G. HEP2 cells were infected with the indicated viruses. At 26 hours post-infection, infected cells were incubated with F, G, or N antibodies, and relative F and G surface levels were determined using ELISA (the F protein is an indicator of viral genomic replication and is shown for normalization purposes; to detect N, cells are detergent-permeabilized). Four F antibodies (provided by J S McEwan) were used to detect F. D25, 14402, and AM14 are specific for pre-fusion F. AM14 only recognizes correctly maximized mature pre-fusion F. 15576 is specific for the postfusion conformation. Abundance of 15576 signal shows that preF\(^{\text{ACCT}}\) is entirely in the pre-fusion conformation. Abundant levels of conformationally correct pre-fusion F were expressed at the surface of vaccine-virus infected cells. As expected, the G protein was detected at similar levels. All F Abs were applied at 0.1 µg/ml. Error bars are standard deviation of the mean from triplicate samples.

FIG. 6 graphically depicts that vaccine candidate RS6-preF\(^{\text{ACCT}}\) secreted high levels of pre-fusion F (this is the codon-optimized preF gene). HEP2 cells were infected with the indicated viruses. At 36 hours post-infection, supernatants of infected cells were harvested and incubated on ELISA plates coated with the anti-tag antibodies for 1 hour. Bound preF\(^{\text{ACCT}}\) was then detected by ELISA using D25 and monoclonal antibody as primary antibodies.

FIG. 7 graphically depicts that preF expressing single cycle RSV induced high levels of anti-RSV antibodies in vivo. 96 well plates were coated with preF-G by infecting HEP-2 cells with RS6-preF\(^{\text{ACCT}}\). At 26 hours post-infection, preF and G proteins were present at the cell surface in conformationally accurate (native) form (as shown in FIG. 5). Pooled sera (n=3, collected at 3 weeks post-boost) from mice vaccinated prime/boost with RS6-preF\(^{\text{ACCT}}\) or RecWT virus were incubated on the coated ELISA plates, and antibody levels were determined using ELISA.

FIG. 8 graphically depicts a schematic overview of examples of different preF-based single cycle RSV vaccines. First panel, RS6V-preF\(^{\text{ACCT}}\); second panel, RS6-preF\(^{\text{ACCT}}\), NS1low; third panel, RS6V-preF\(^{\text{ACCT}}\)NS2low; fourth panel, RS1V-preF\(^{\text{ACCT}}\)-NS1low; fifth panel, RS2V-preF\(^{\text{ACCT}}\), NS2low.

FIG. 9 graphically depicts that different preF RSV vaccines induced high levels of preF and G protein at the cell surface. HEP-2 cells were infected with viruses RS6-preF\(^{\text{ACCT}}\) and RS1V-preF\(^{\text{ACCT}}\)-NS1low. At 26 hours post-infection, infected cells were incubated with anti-preF and anti-G antibodies, which were subsequently detected by standard ELISA method. Anti-N antibody was also used as an indicator for viral genomic replication (N encapsulates the viral genome), and preF and G antibody levels were determined without and with N level-based normalization.

FIG. 10 graphically depicts that different preF RSV vaccines induced high levels of preF-specific and G-specific antibodies in vivo. 96 well plates were coated with preF-G or preF alone as follows: HEP-2 cells were infected with RS6-preF\(^{\text{ACCT}}\) or RS6-preF\(^{\text{ACCT}}\)-AG (G gene removed). At 26 hours post-infection, either preF-G or preF alone were present at the cell surface in conformationally accurate (native) form (as shown in FIG. 5). Pooled sera (n=3, collected at 3 weeks post-boost) from mice vaccinated prime/boost with RS6-preF\(^{\text{ACCT}}\) and RS1V-preF\(^{\text{ACCT}}\)-NS1low were incubated on the coated ELISA plates (preF-G on the left, preF alone on the right), and antibody levels were determined using standard ELISA method. Anti-G Ab L9 was used to verify the absence of G protein in the preF-alone ELISA.

FIG. 11 graphically depicts that two distinct preF RSV vaccines induced higher neutralizing antibody activity than a wildtype virus, despite being designed as safe, single-cycle vaccines. Three-fold dilutions of pooled mice sera (3 mice per pool; sera harvested 3 weeks post-boost) were incubated with 500 PFU of virus RSV-AG-HRP, which lacks the G protein (allowing detection of F-specific neutralization) and contains the HRP gene for detection. After a one hour incubation, virus-antibody suspensions were incubated on HEP-2 cells for 1.5 hours. Inoculum was removed and cells incubated for a total of 48 hours post-infection (hpi). At 48 hpi, medium was replaced with standard (TM1) ELISA substrate, and OD\(_{405}\) was determined after 30 minutes as a measure of virus replication.

FIG. 12 graphically depicts that the preF RSV vaccine RS6-preF\(^{\text{ACCT}}\) (codon-optimized PreF) induced higher neutralizing antibody activity than a wildtype virus, despite being designed as a safe, single-cycle vaccine. Three-fold dilutions of pooled mice sera (3 mice per pool; sera harvested 3 weeks post-boost) were incubated with 250 PFU of virus RSV-AG-HRP, which lacks the G protein (allowing detection of F-specific neutralization) and contains the HRP gene for detection. After a one hour incubation, virus-antibody suspensions were incubated on HEP-2 cells for 1.5 hours. Inoculum was removed and cells incubated for a total of 48 hours post-infection (hpi). At 48 hpi, medium was replaced with standard (TM1) ELISA substrate, and OD\(_{405}\) was determined after 30 minutes as a measure of virus replication.

DETAILED DESCRIPTION

Before explaining at least one embodiment of the inventive concept(s) in detail by way of exemplary language and results, it is to be understood that the inventive concept(s) is not limited in its application to the details of construction and the arrangement of the components set forth in the following description. The inventive concept(s) is capable of other embodiments or of being practiced or carried out in various ways. As such, the language used herein is intended to be given the broadest possible scope and meaning, and the embodiments are meant to be exemplary—not exhaustive.
Also, it is to be understood that the phraseology and terminology employed herein is for the purpose of description and should not be regarded as limiting.

Unless otherwise defined herein, scientific and technical terms used in connection with the presently disclosed inventive concept(s) shall have the meanings that are commonly understood by those of ordinary skill in the art. Further, unless otherwise required by context, singular terms shall include plurals, and plural terms shall include the singular. The foregoing procedures and techniques are generally performed according to conventional methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification. The nomenclature utilized in connection with, and the laboratory procedures and techniques of, analytical chemistry, synthetic organic chemistry, and medicinal and pharmaceutical chemistry described herein are those well-known and commonly used in the art. Standard techniques are used for chemical syntheses and chemical analyses.

All patents, published patent applications, and non-patent publications mentioned in the specification are indicative of the level of skill of those skilled in the art to which this presently disclosed inventive concept(s) pertains. All patents, published patent applications, and non-patent publications referenced in any portion of this application are herein expressly incorporated by reference in their entirety to the same extent as if each individual patent or publication was specifically and individually indicated to be incorporated by reference.

All of the compositions and/or methods disclosed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of the inventive concept(s) have been described in terms of particular embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and/or methods and in the steps or in the sequence of steps of the methods described herein without departing from the concept, spirit, and scope of the inventive concept(s). All such similar substitutions and modifications apparent to those skilled in the art are deemed to be within the spirit, scope, and concept of the inventive concept(s) as defined by the appended claims.

As utilized in accordance with the present disclosure, the following terms, unless otherwise indicated, shall be understood to have the following meanings:

The use of the term “a” or “an” when used in conjunction with the term “comprising” in the claims and/or the specification may mean “one,” but it is also consistent with the meaning of “one or more,” “at least one,” and “one or more than one.” As such, the terms “a,” “an,” and “the” include plural referents unless the context clearly indicates otherwise. Thus, for example, reference to “a compound” may refer to one or more compounds, two or more compounds, three or more compounds, four or more compounds, or greater numbers of compounds. The term “plurality” refers to “two or more.”

The use of the term “at least one” will be understood to include one as well as any quantity more than one, including but not limited to, 2, 3, 4, 5, 10, 15, 20, 30, 40, 50, 100, etc. The term “at least one” may extend up to 100 or 1000 or more, depending on the term to which it is attached; in addition, the quantities of 100/1000 are not to be considered limiting, as higher limits may also produce satisfactory results. In addition, the use of the term “at least one of X, Y, and Z” will be understood to include X alone, Y alone, and Z alone, as well as any combination of X, Y, and Z. The use of ordinal number terminology (i.e., “first,” “second,” “third,” “fourth,” etc.) is solely for the purpose of differentiating between two or more items and is not meant to imply any sequence or order or importance to one item over another or any order of addition, for example.

The use of the term “or” in the claims is used to mean an inclusive “and/or” unless explicitly indicated to refer to alternatives only or unless the alternatives are mutually exclusive. For example, a condition “A or B” is satisfied by any of the following: A is true (or present) and B is false (or not present), A is false (or not present) and B is true (or present), and both A and B are true (or present).

As used herein, any reference to “one embodiment,” “an embodiment,” “some embodiments,” “one example,” “for example,” or “an example” means that a particular element, feature, structure, or characteristic described in connection with the embodiment is included in at least one embodiment. The appearance of the phrase “in some embodiments” or “one example” in various places in the specification is not necessarily all referring to the same embodiment, for example. Further, all references to one or more embodiments or examples are to be construed as non-limiting to the claims.

Throughout this application, the term “about” is used to indicate that a value includes the inherent variation of error for a composition/apparatus/device, the method being employed to determine the value, or the variation that exists among the study subjects. For example, but not by way of limitation, when the term “about” is utilized, the designated value may vary by plus or minus twenty percent, or fifteen percent, or twelve percent, or eleven percent, or ten percent, or nine percent, or eight percent, or seven percent, or six percent, or five percent, or four percent, or three percent, or two percent, or one percent from the specified value, as such variations are appropriate to perform the disclosed methods and as understood by persons having ordinary skill in the art.

As used in this specification and claim(s), the words “comprising” (and any form of comprising, such as “comprise” and “comprises”), “including” (and any form of including, such as “includes” and “include”), “containing” (and any form of containing, such as “contains” and “contain”) are inclusive or open-ended and do not exclude additional, unreferenced elements or method steps.

The term “or combinations thereof” as used herein refers to all permutations and combinations of the listed items preceding the term. For example, “A, B, C, or combinations thereof” is intended to include at least one of: A, B, C, AB, AC, BC, or ABC, and if order is important in a particular context, also BA, CA, CB, CBA, BCA, ACB, BAC, or CAB. Continuing with this example, expressly included are combinations that contain repeats of one or more item or term, such as BB, AAA, AAB, BBC, AAAABCCCC, CBBAAA, CABABB, and so forth. The skilled artisan will understand that typically there is no limit on the number of items or terms in any combination, unless otherwise apparent from the context.

As used herein, the term “substantially” means that the subsequently described event or circumstance completely occurs or that the subsequently described event or circumstance occurs to a great extent or degree. For example, when associated with a particular event or circumstance, the term “substantially” means that the subsequently described event or circumstance occurs at least 80% of the time, or at least 85% of the time, or at least 90% of the time, or at least 95% of the time. For example, the term “substantially adjacent” may mean that two items are 100% adjacent to one another,
or that the two items are within close proximity to one another but not 100% adjacent to one another, or that a portion of one of the two items is not 100% adjacent to the other item but is within close proximity to the other item.

The term “polypeptide” as used herein will be understood to refer to a polymer of amino acids. The polymer may include D-, L-, or artificial variants of amino acids. In addition, the term “polypeptide” will be understood to include peptides, proteins, and glycoproteins.

The term “polynucleotide” as used herein will be understood to refer to a polymer of two or more nucleotides. Nucleotides, as used herein, will be understood to include deoxyribose nucleotides and/or ribose nucleotides, as well as artificial variants thereof. The term polynucleotide also includes single-stranded and double-stranded molecules.

The terms “analog” or “variant” as used herein will be understood to refer to a variation of the normal or standard form or the wild-type form of molecules. For polypeptides or polynucleotides, an analog may be a variant (polymorphism), a mutant, and/or a naturally or artificially chemically modified version of the wild-type polypeptide (including combinations of the above). Such analogs may have higher, full, intermediate, or lower activity than the normal form of the molecule, or no activity at all. Alternatively and/or in addition thereto, for a chemical analog may be any structure that has the desired functionalities (including alterations or substitutions in the core moiety), even if comprised of different atoms or isomeric arrangements.

As used herein, the phrases “associated with” and “coupled to” include both direct association/binding of two moieties to one another as well as indirect association/binding of two moieties to one another. Non-limiting examples of associations/couplings include covalent binding of one moiety to another moiety either by a direct bond or through a spacer group, non-covalent binding of one moiety to another moiety either directly or by means of specific binding pair members bound to the moieties, incorporation of one moiety into another moiety such as by dissolving one moiety in another moiety or by synthesis, and coating one moiety on another moiety, for example.

As used herein, “substantially pure” means an object species is the predominant species present (i.e., on a molar basis it is more abundant than any other individual species in the composition), and preferably a substantially purified fraction is a composition wherein the object species comprises at least about 50 percent (on a molar basis) of all macroscopic species present. Generally, a substantially pure composition will comprise more than about 80 percent of all macroscopic species present in the composition, more preferably more than about 85%, 90%, 95%, and 99%. Most preferably, the object species is purified to essential homogeneity (contaminant species cannot be detected in the composition by conventional detection methods) wherein the composition consists essentially of a single macroscopic species.

The term “pharmaceutically acceptable” refers to compounds and compositions which are suitable for administration to humans and/or animals without undue adverse side effects such as (but not limited to) toxicity, irritation, and/or allergic response commensurate with a reasonable benefit/risk ratio.

The term “patient” as used herein includes human and veterinary subjects. “Mammal” for purposes of treatment refers to any animal classified as a mammal, including (but not limited to) humans, domestic and farm animals, nonhuman primates, and any other animal that has mammary tissue.

The term “child” is meant to refer to a human individual who would be recognized by one of skill in the art as an infant, toddler, etc., or an individual less than about 18 years of age, usually less than about 16 years of age, usually less than about 14 years of age, or even less (e.g., from newborn to about 2 years of age, or about 2, about 2.5, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 11, or about 12 years of age). The term “elderly” generally refers to a human individual whose age is greater than about 50 years of age, usually greater than about 55 years of age, frequently greater than about 60 years of age or more (e.g., about 65 years of age and upwards).

The term “treatment” refers to both therapeutic treatment and prophylactic or preventative measures. Those in need of treatment include, but are not limited to, individuals already having a particular condition/disease/infection as well as individuals who are at risk of acquiring a particular condition/disease/infection (e.g., those needing prophylactic/preventative measures). The term “treating” refers to administering an agent to a patient for therapeutic and/or prophylactic/preventative purposes.

A “therapeutic composition” or “pharmaceutical composition” refers to an agent that may be administered in vivo to bring about a therapeutic and/or prophylactic/preventative effect.

Administering a therapeutically effective amount or prophylactically effective amount is intended to provide a therapeutic benefit in the treatment, prevention, and/or management of a disease, condition, and/or infection. The specific amount that is therapeutically effective can be readily determined by the ordinary medical practitioner, and may vary depending on factors known in the art, such as (but not limited to) the type of condition/disease/infection, the patient’s history and age, the stage of the condition/disease/infection, and the co-administration of other agents.

The term “effective amount” refers to an amount of a biologically active molecule or conjugate or derivative thereof sufficient to exhibit a detectable therapeutic effect without undue adverse side effects (such as (but not limited to) toxicity, irritation, and allergic response) commensurate with a reasonable benefit/risk ratio when used in the manner of the inventive concept(s). The therapeutic effect may include, for example but not by way of limitation, preventing, inhibiting, or reducing the occurrence of infection by or growth of microbes and/or opportunistic infections. The effective amount for a subject will depend upon the type of subject, the subject’s size and health, the nature and severity of the condition/disease/infection to be treated, the method of administration, the duration of treatment, the nature of concurrent therapy (if any), the specific formulations employed, and the like. Thus, it is not possible to specify an exact effective amount in advance. However, the effective amount for a given situation can be determined by one of ordinary skill in the art using routine experimentation based on the information provided herein.

As used herein, the term “concurrent therapy” is used interchangeably with the terms “combination therapy” and “adjunct therapy,” and will be understood to mean that the patient in need of treatment is treated or given another drug for the disease/infection in conjunction with the pharmaceutical compositions of the present disclosure. This concurrent therapy can be sequential therapy, where the patient is treated first with one pharmaceutical composition and then the other pharmaceutical composition, or the two pharmaceutical compositions are given simultaneously.

The terms “administration” and “administering,” as used herein, will be understood to include all routes of administration known in the art, including but not limited to, oral,
topical, transdermal, parenteral, subcutaneous, intranasal, mucosal, intramuscular, intraperitoneal, intraviteal, and intravenous routes, and including both local and systemic applications. In addition, the compositions of the present disclosure (and/or the methods of administration of same) may be designed to provide delayed, controlled, or sustained release using formulation techniques which are well known in the art.

Turning now to the inventive concept(s), certain non-limiting embodiments of the present disclosure are directed to a recombinant, live, attenuated virus of the Pneumoviridae family. The recombinant, live, attenuated virus includes a baculovirus GP64 envelope glycoprotein or variant or fragment thereof and a polynucleotide encoding a respiratory syncytial virus (RSV) F protein variant or fragment thereof. The baculovirus GP64 envelope glycoprotein or fragment thereof is capable of mediating entry of the recombinant virus into a mammalian cell. The respiratory syncytial virus (RSV) F protein variant or fragment thereof includes at least one amino acid substitution when compared to a native RSV F protein, wherein the at least one amino acid substitution stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation.

In certain non-limiting embodiments, the recombinant, live, attenuated virus is isolated from the cell in which it is produced.

In certain non-limiting embodiments, the recombinant, live, attenuated virus is further defined as a recombinant respiratory syncytial virus (RSV).

In certain non-limiting embodiments, the recombinant, live, attenuated virus is capable of infecting a cell in a mammal but cannot transmit from said cell to another cell in the mammal.

In certain non-limiting embodiments, the recombinant, live, attenuated virus is further defined as an enveloped recombinant, live, attenuated virus.

Also, in certain non-limiting embodiments, the recombinant, live, attenuated virus maintains infective stability when stored at above 0° C. for at least 3.5 days.

Any baculovirus GP64 envelope glycoprotein, variant thereof, or fragment thereof known in the art or otherwise contemplated herein may be utilized in accordance with the present disclosure, so long as the protein/variant/fragment is capable of mediating entry of the recombinant virus into a mammalian cell. In certain particular (but non-limiting) embodiments, the baculovirus GP64 envelope glycoprotein or variant or fragment thereof comprises an ectodomain of the baculovirus GP64 envelope glycoprotein, a transmembrane domain of the baculovirus GP64 envelope glycoprotein, and/or a heterologous cytoplasmic tail (such as, but not limited to, a polypeptide from the F protein, such as, but not limited to, a 12 amino acid polypeptide). Non-limiting examples of GP64 proteins/variants/fragments that may be utilized in accordance with the present disclosure are disclosed in US Patent Publication Application No. 2007/0104734, published May 10, 2007 to Oomens et al. and U.S. Pat. No. 7,588,770, issued Sep. 15, 2009 to Oomens et al., as well as Oomens et al. (Journal of Virology (2004) 78:9064-9072); the entire contents of each of the above references are hereby expressly incorporated herein by reference. One particular (but non-limiting) example of a GP64 glycoprotein variant disclosed in the above references that may be utilized in accordance with the present disclosure is GP64 \( \text{GP}^{\text{F}_{-1}} \), in which the 7-amino acid cytoplasmic tail domain of GP64 was replaced by the 12 C-terminal amino acids of the HRSV F protein; the amino acid sequence of GP64 is represented by SEQ ID NO:15, and the nucleotide sequence encoding same is represented by SEQ ID NO:16.

In addition, the baculovirus GP64 envelope glycoprotein or variant or fragment thereof may not be directly encoded by the virus but rather is obtained from the cell line from which the virus is produced. Alternatively, the GP64 glycoprotein or variant or fragment thereof may be encoded by the virus.

Also, the recombinant, live, attenuated virus may further encode at least one other protein normally encoded by the virus' wild type genome, or may further encode at least one variant or fragment thereof. For example (but not by way of limitation), the virus may further encode at least one of RSV NS1 protein or a variant or fragment thereof; NS2 protein or a variant or fragment thereof; N protein or a variant or fragment thereof; P protein or a variant or fragment thereof; M protein or a variant or fragment thereof; M-2 protein or a variant or fragment thereof; L protein or a variant or fragment thereof; or any combination thereof. One particular (but non-limiting) variant or fragment of a genomic protein that may be utilized is the secreted G protein (known as Gmem); the amino acid sequence of Gmem is represented by SEQ ID NO:13, and the nucleotide sequence encoding same is represented by SEQ ID NO:14. Alternatively, the wild type RSV G protein may be present in any of the recombinant, live, attenuated viruses of the present disclosure; the gene encoding the wild type RSV G protein is represented by SEQ ID NO:17, while a codon-optimized sequence encoding the wild type RSV G protein is represented by SEQ ID NO:18.

In certain embodiments, the virus may be further defined as lacking expression of at least one virulence factor encoded by the wild type virus, such as (but not limited to), the NS1 or NS2 protein or Gmem.

Any RSV F protein variant or fragment thereof known in the art or otherwise contemplated herein may be utilized in accordance with the present disclosure, so long as the RSV F protein variant or fragment thereof includes at least one amino acid substitution compared to a native RSV F protein that stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation. Any amino acid substitution(s) capable of stabilizing the RSV F protein variant/fragment in the pre-fusion confirmation may be utilized in accordance with the present disclosure. Particular (but non-limiting) examples of RSV F protein variants or fragments thereof that may be utilized in accordance with the present disclosure include RSV F protein variants or fragments thereof that include at least one, at least two, at least three, or all four of the amino acid substitutions S155C, 5190F, V207L, and S290C when compared to the native RSV F protein sequence, as represented by SEQ ID NO:1. For example (but not by way of limitation), the RSV F protein variant or fragment thereof can comprise an amino acid sequence represented by at least one of SEQ ID Nos:2-4 (see Table 1 and FIG. 1).

In certain non-limiting embodiments, the RSV F protein variant or fragment thereof is absent a portion or all of a cytoplasmic tail and/or a portion or all of a transmembrane domain of the native RSV F protein. Alternatively, the RSV F protein variant or fragment thereof may include a portion or all of the cytoplasmic tail and/or a portion or all of the transmembrane domain of the native RSV F protein. In one particular (but non-limiting) embodiment, the transmembrane domain approximately corresponds to residues 525-550 of SEQ ID NO:1, while the cytoplasmic tail approximately corresponds to residues 554-574 of SEQ ID NO:1.
Alternatively (and/or in addition thereto), the RSV F protein variant or fragment thereof further comprises at least one epitope tag. One non-limiting example of an epitope tag that may be utilized in accordance with the present disclosure is the AcV5 epitope tag. The amino acid sequence of the AcV5 epitope tag is represented by SEQ ID NO:11, and the nucleotide sequence of this tag is represented by SEQ ID NO:12.

Alternatively (and/or in addition thereto), the RSV F protein variant or fragment thereof may further include a detectable marker.

As stated herein above, any amino acid substitution(s) capable of stabilizing the RSV F protein variant/fragment in the pre-fusion confirmation may be utilized in accordance with the present disclosure. Non-limiting examples of RSV F protein variants or fragments thereof (that contain one or more amino acid substitution(s) capable of stabilizing the RSV F protein variant/fragment in the pre-fusion confirmation) are disclosed in US Patent Application Publication Nos. US 2015/0030622, US 2016/001972, and US 2016/0046675 (all incorporated supra); McEllan et al. (Science (2013) 342:592-598); Krarup et al. (Nature Communications (2015) 6:8143 (Pages 1-12); and Joyce et al. (Nature Structural and Molecular Biology (2016) 23:811-822); the entire contents of these references being expressly incorporated herein by reference.

Any nucleotide encoding any of the RSV F protein variants or fragments thereof may be utilized in accordance with the present disclosure. In certain non-limiting embodiments, the polynucleotide sequence corresponding to the wild type RSV F protein sequence (except for the codons encoding the amino acid substitution(s)). Alternatively, the polynucleotide sequence may be codon-optimized to increase expression thereof in a host cell. For example, as shown in Table 1, SEQ ID NOS:8, 9, and 10 contain polynucleotide sequences that encode the variants of SEQ ID NOS:2, 3, and 4, respectively, and are identical to the corresponding portion of the nucleotide sequence of the wild type F protein sequence, with the exception of the codons encoding for the amino acid substitutions. Alternatively, SEQ ID NOS:5, 6, and 7 also encode the variants of SEQ ID NOS:2, 3, and 4, respectively, but these polynucleotides have been codon-optimized to increase the expression thereof in a host cell.

**TABLE 1**

<table>
<thead>
<tr>
<th>RSV F Protein Variant</th>
<th>Nucleotide Sequence Based on WT RSV F Sequence</th>
<th>Nucleotide Sequence Based on Codon-Optimized RSV F Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>prepSEC</td>
<td>SEQ ID NO: 2</td>
<td>SEQ ID NO: 5</td>
</tr>
<tr>
<td>prepSEC</td>
<td>SEQ ID NO: 3</td>
<td>SEQ ID NO: 6</td>
</tr>
<tr>
<td>prepSEC+Corr</td>
<td>SEQ ID NO: 4</td>
<td>SEQ ID NO: 7</td>
</tr>
</tbody>
</table>

*See FIG. 1 for the structures of each of the RSV F protein variants.*

In certain non-limiting embodiments, the recombinant, live, attenuated virus further comprises the expressed RSV F protein variant or fragment thereof (as opposed to simply including the polynucleotide sequence encoding same).

Certain non-limiting embodiments of the present disclosure are also directed to an isolated immunogenic composition comprising any of the viruses described or otherwise contemplated herein.

Further non-limiting embodiments of the present disclosure are directed to at least one cell that is capable of producing any of the recombinant, live, attenuated viruses described or otherwise contemplated herein. The cell(s) includes: (i) at least one polynucleotide encoding a baculovirus GP64 envelope glycoprotein or variant or fragment thereof; and (ii) at least one polynucleotide encoding at least a portion of a genome of an infection-attenuated virus of the Pseudomonas family, wherein the genome comprises a gene encoding an RSV F protein variant or fragment thereof that comprises at least one amino acid substitution compared to a native RSV F protein, wherein the at least one amino acid substitution stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation. In certain non-limiting embodiments, the RSV genome includes other additions or modifications thereto.

Any cell type capable of producing the recombinant, live, attenuated viruses and capable of functioning as described or otherwise contemplated herein falls within the scope of the present disclosure. In certain non-limiting embodiments, the cell is a mammalian cell. In certain non-limiting embodiments, the cell is a Vbace cell, which is a Vero cell stably transfected with the baculovirus GP64 protein carrying a portion of the cytoplasmic tail of RSV F protein (i.e., GP64C) (as disclosed in Oomens et al. (2004); US Patent Application Publication No. 2007/0104734; and U.S. Pat. No. 7,588,770; incorporated supra). In addition, the present disclosure also includes modified versions of any of the above cell lines. For example (but not by way of limitation), a Vbace cell line can be modified to include additional genetic modifications to GP64C, including one or more modifications to the GP64 portion of the sequence and/or one or more modifications to the F cytoplasmic tail portion. Alternatively, other cell lines may be modified to express GP64 or a variant thereof (such as, but not limited to, GP64C; a modified form thereof, or another modified form of GP64). These modified cell lines may be produced to improve on the growth characteristics of the cell line and/or to improve on the cell line's ability to produce virus, thereby enhancing production of the compositions of the present disclosure.

In still yet another aspect, mammalian cells or mammals are provided which include a recombinant virus as described or otherwise contemplated herein, or which include polynucleotide(s) that encode all of the various components of the recombinant virus, as described or otherwise contemplated herein.

In a particular (but non-limiting) embodiment, a mammalian cell of the present disclosure includes an expression cassette encoding a heterologous envelope protein comprising an ectodomain of a baculovirus transmembrane protein, and one or more expression vectors comprising or expressing the genome of an infection-defective or infection-attenuated mammalian virus as known in the art or as described or otherwise contemplated herein. The expression cassette can be stably or transiently transfected or transduced into the mammalian cell. In one example, the expression cassette is integrated into a chromosome of the mammalian cell. In another example, the mammalian cell is a Vero cell.

The infection-defective or -attenuated mammalian virus, when assembled in the mammalian cell, incorporates the heterologous envelope protein which affords the virus with improved infectivity and/or stability. In one example, the mammalian virus is a recombinant RSV, and the heterologous envelope protein comprises an ectodomain of baculovirus envelope GP64 protein. In another example, the recombinant RSV lacks one or more functional transmembrane proteins, such as SH, G, or F proteins. The recombi-
nant RSV also includes a pre-fusion F protein variant as described or otherwise contemplated herein.

Furthermore, certain non-limiting embodiments of the present disclosure are directed to mammalian cells, such as (but not limited to) Vero cells, that are stably transfected or transduced with at least one expression cassette encoding a recombinant viral envelope protein (or variant or fragment thereof) and encoding a pre-fusion F protein variant. The viral envelope protein includes an ectodomain of a baculovirus transmembrane protein (e.g., the GP64 protein). The pre-fusion F protein variant is as described or otherwise contemplated herein. These recombinant RSVs are attenuated for cell-to-cell transmission.

Recombinant mammalian or vertebrate viruses other than pneumoviruses can be similarly prepared using the present disclosure. These viruses may have all of the advantageous properties possessed by the recombinant pneumoviruses that are disclosed or otherwise contemplated herein. For instance, these viruses can have improved stability of infectivity as compared to their wild-type counterparts. In addition, these viruses can be temperature sensitive and infectious but incapable of spreading between host cells. In one embodiment, these viruses include heterologous envelope proteins that comprise the ectodomain of a baculovirus transmembrane protein, such as (but not limited to) the GP64 protein or its functional equivalents, and a pre-fusion F protein variant. In another embodiment, these viruses are not recombinant lentiviruses, such as (but not limited to) those described in Kumar, et al. (Human Gene Therapy (2003) 14:67-77) and Ojala, et al. (Biochem. Biophys. Res. Commun. (2001) 284:777-784), the entire contents of each of which are hereby expressly incorporated herein by reference.

While certain non-limiting embodiments of the present disclosure are directed to viral production whereby the baculovirus GP64 protein/variant/fragment is supplied in trans to RSV from the cell line in which the virus is produced, it should be understood that the scope of the present disclosure also includes modifying the RSV genome to directly contain the gene encoding the GP64 protein/variant/fragment. In this manner, the RSV genome encodes both: (i) any of the RSV F protein variants/fragments described or otherwise contemplated herein; and (ii) any of the GP64 proteins/variants/fragments described or otherwise contemplated herein. Having GP64 encoded in the viral genome ensures strong GP64 expression levels, which improves virus production and virus temperature stability. When the GP64 protein is provided in cis, the RSV genome will need to be further modified so as to provide a self-limited safety component thereto (i.e., by inactivating one or more essential viral components). One non-limiting example of such a modification is to replace the M gene with that of the tet-transactivator gene; the resulting vaccine virus is then amplified in the laboratory by growing in M-expressing cells, whereby M is expressed via tet-responsive promoters.

Another non-limiting example of such a modification is to replace the M gene with any suitable non-RSV gene, whereby the resulting vaccine virus is then amplified in the laboratory by growing in M-expressing cells, whereby M is expressed via inducible or constitutive promoters.

Further non-limiting embodiments of the present disclosure are directed to a pharmaceutical composition that includes a therapeutically effective amount of any of the recombinant, live, attenuated viruses described in detail herein above or otherwise contemplated herein. Alternatively and/or in addition thereto, the pharmaceutical composition may include any of the polynucleotides described or otherwise contemplated herein. In certain non-limiting embodiments, the pharmaceutical composition is capable of eliciting an immune response against the virus or a component thereof in a mammal. In particular (but non-limiting) embodiments, the therapeutically effective amount of the recombinant, live, attenuated virus is further defined as an amount sufficient to induce an immune response protective against RSV infection. Thus, in a particular (but non-limiting) embodiment, the pharmaceutical composition may be an immunogenic composition, such as (but not limited to) a vaccine.

The pharmaceutical compositions or formulations disclosed or otherwise contemplated herein include one or more attenuated viruses as described herein, each of which is substantially purified and/or isolated, except that one or more of such viruses may be included in a single composition. In certain non-limiting embodiments, the pharmaceutical compositions also include a pharmaceutically acceptable carrier or excipient. Any carriers or excipients known in the art may be utilized in accordance with the present disclosure. For example (but not by way of limitation), a physiological compatible carrier (e.g., saline) that is compatible with maintaining the infectivity of the virus when administered (i.e., the viruses that are initially administered are capable of infecting one or more host cells), and compatible with the desired mode of administration, may be utilized as the pharmaceutically acceptable carrier in accordance with the present disclosure. In addition, the active ingredients may be mixed with excipients which are pharmaceutically acceptable and compatible with the active ingredients. Suitable excipients include, for example but not by way of limitation, water, saline, dextrose, glycerol, ethanol, and the like, or any combination thereof.

The preparation of such compositions for use as immunogenic compositions, such as (but not limited to) vaccines, is well known to those of skill in the art. Typically, such compositions are prepared either as liquid solutions or suspensions; however, solid forms such as (but not limited to) tablets, pills, powders, and the like are also contemplated. Solid forms suitable for solution in, or suspension in, liquids prior to administration may also be prepared. The preparation may also be emulsified. In addition, the pharmaceutical compositions disclosed or otherwise contemplated herein may contain minor amounts of auxiliary substances, such as (but not limited to) wetting or emulsifying agents, pH buffering agents, and the like, as well as any combination thereof. If it is desired to administer an oral form of the pharmaceutical composition, one or more of various thickeners, flavorings, dyes, and emulsifiers, dispersing aids, binders, or the like, as well as any combination thereof, may be added. The pharmaceutical compositions of the present disclosure may contain any such additional ingredients so as to provide the composition in a form suitable for administration.

In addition, in certain non-limiting embodiments, the pharmaceutical composition contains at least one adjuvant. Suitable adjuvants are well known to those skilled in the art and include, without limitation, aluminum phosphate; at least one saponin complexed to at least one membrane protein antigen to produce immune stimulating complex(es) (ISCOMs); at least one pluron polymer with mineral oil; killed mycobacteria in mineral oil; Freund’s complete adjuvant; at least one bacterial product, such as (but not limited to) muramyl dipeptide (MDP) and lipopolysaccharide (LPS); monophosphoryl lipid A; QS 21; and polysphosphazene, as well as any component or derivative thereof; and as well as any combination thereof.
The recombinant, live, attenuated virus may be present in the pharmaceutical composition at any percentage of concentration that allows the virus to function as described or as otherwise contemplated herein. For example (but not by way of limitation), the virus may be present in a sufficient amount to function as an immunogenic composition. In certain particular (but non-limiting) embodiments, the recombinant, live, attenuated virus is present in the pharmaceutical composition at a percent concentration of about 0.001%, about 0.005%, about 0.01%, about 0.05%, about 0.1%, about 0.5%, about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 95%, and about 99%. In addition, the scope of the presently disclosed also includes the presence of the virus in the pharmaceutical composition at any percent concentration that falls within any range formed from the combination of two values listed above (for example, a range of from about 1% to about 99%, a range of from about 2% to about 80%, a range of from about 3% to about 60%, a range of from about 10% to about 95%, a range of from about 40% to about 75%, etc.).

Likewise, a pharmaceutically acceptable carrier, excipient, and/or adjuvant may be present in the pharmaceutical composition at any percentage of concentration that allows the carrier/excipient/adjuvant to function as described or as otherwise contemplated herein. In certain particular (but non-limiting) embodiments, each of the pharmaceutically acceptable carrier, excipient, and adjuvant is present in the pharmaceutical composition at a percent concentration of about 0.001%, about 0.005%, about 0.01%, about 0.05%, about 0.1%, about 0.5%, about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 95%, and about 99%. In addition, the scope of the presently disclosure also includes the presence of each of the pharmaceutically acceptable carrier, excipient, and adjuvant in the pharmaceutical composition at any percent concentration that falls within any range formed from the combination of two values listed above (for example, a range of from about 1% to about 99%, a range of from about 2% to about 80%, a range of from about 3% to about 60%, a range of from about 10% to about 95%, a range of from about 40% to about 75%, etc.).

The pharmaceutical compositions of the present disclosure may be administered by any of the many suitable means described herein and/or which are well known to those of skill in the art, including but not limited to: by injection, inhalation, oral, intravaginal, intranasal, rectal, or intradermal administration; by ingestion of a food or probiotic product containing the virus; by topical administration, such as (but not limited to) eye drops, sprays, etc.; and the like. In one instance, the administration will be carried out by using an implant. In particular (but non-limiting) embodiments, the mode of administration is by injection and/or inhalation. One or more than one route of administration can be employed either simultaneously or partially or wholly sequentially, i.e., prime boost vaccine regimens are also contemplated. Such prime boost vaccine regimens typically involve repeated vaccine administration at preselected intervals, such as (but not limited to) at 1 month or 6 weeks of age then at 6 months, 1 year, and yearly thereafter, or at longer intervals, e.g., every 5 or 10 years, etc. Those of skill in the art are well acquainted with the planning, implementation, and assessment of such vaccine strategies, and therefore no further discussion thereof is required.

The pharmaceutical compositions may be administered in conjunction with other treatment modalities. In some embodiments, such modalities may include (but are not limited to) various substances that boost the immune system, various chemotherapeutic agents, vitamins, anti-allergy agents, anti-inflammatory agents, etc. In other embodiments, other antigenic agents (e.g., other vaccines or vaccine components), may be advantageously administered or co-administered with the pharmaceutical compositions disclosed or otherwise contemplated herein. For example, in some cases it may be desirable to combine any of the recombinant virus pharmaceutical compositions disclosed or otherwise contemplated herein with known vaccines which induce protective responses to other agents, particularly other childhood viruses or other infectious agents. The other vaccines may also be live attenuated virus vaccines, but this need not always be the case; such vaccines may be inactivated virus vaccines or vaccines against other etiological agents (e.g., bacteria). When multiple immunogenic compositions/vaccines are to be administered together, the immunogenic compositions/vaccine agents may be combined in a single pharmaceutical composition. Alternatively (and/or in addition thereto), the multiple immunogenic compositions/vaccines may be administered separately but over a short time interval, e.g., at a single visit at a doctor's office or clinic, etc.

In addition, the attenuated viruses disclosed or otherwise contemplated herein may also be further genetically engineered to contain and express genes encoding other antigens and/or agents of interest. The other agents of interest may, for example (but not by way of limitation), include a foreign epitope or another "tag" or "marker" of heterologous or foreign genetic material. Such agents are useful, for example, for distinguishing between wildtype and vaccine viral strains, such as (but not limited to) in the laboratory, in nature, in a host, etc. Basically, a genetically engineered recombinant virus disclosed or otherwise contemplated herein would carry the tag, but the wildtype virus would not. This technique can also be used to distinguish between viral vaccine strains, permitting the introduction of unique genetic tags into different batches or different iterations of recombinant virus strains, to detect pirated formulations, etc.

In addition, the genetically engineered viruses may contain and express detectable markers (e.g., labeling or reporter groups such as (but not limited to) various peptides and/or proteins), for the purpose of tracing or visualizing the location of the viruses, cells infected by the viruses, or proteins translated from the viral genome; or for quantitating viruses or cells infected by virus, etc. Exemplary detectable markers include, but are not limited to: various fluorescent entities such as green fluorescent protein (GFP), blue, cyan, etc. fluorescent protein, and various derivatives thereof; other fluorescent proteins such as (but not limited to) dsRed, eqFP611, Dronpa, TagRFPs, KFP, EosFP, Dendra, IrisFP, etc.; other similar molecules known in the art; and any derivatives or combinations thereof.

Yet further non-limiting embodiments of the present disclosure are directed to a method of producing any of the recombinant, live, attenuated viruses described or otherwise contemplated herein. In one non-limiting embodiment of the method, a cell line is provided that expresses a baculovirus GP64 envelope glycoprotein or variant or fragment thereof; the cell line is also transfected with at least one polynucleotide encoding RSV virus, wherein the RSV virus comprises...
an RSV F protein variant or fragment thereof that comprises at least one amino acid substitution compared to a native RSV F protein, wherein the at least one amino acid substitution stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation. In addition, the cell line is cultured under conditions that allow for production of the recombinant, live, attenuated virus. In certain particular (but non-limiting) embodiments, the recombinant, live, attenuated virus is isolated away from the cultured cells. In a particular (but non-limiting) embodiment, the recombinant, live, attenuated virus is substantially purified.

In certain particular (but non-limiting) embodiments, the method includes: (i) recovering recombinant, live, attenuated virus comprising a polynucleotide encoding a respiratory syncytial virus (RSV) F protein variant or fragment thereof from cDNA using reverse genetics in the presence of a baculovirus GP64 envelope glycoprotein or variant or fragment thereof, wherein the RSV F protein variant or fragment thereof comprises at least one amino acid substitution compared to a native RSV F protein, wherein the at least one amino acid substitution stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation; and (ii) amplifying the attenuated virus in a cell line expressing the baculovirus GP64 envelope glycoprotein or variant or fragment thereof.

Yet further non-limiting embodiments of the present disclosure are directed to a use of any of the recombinant, live, attenuated viruses disclosed or otherwise contemplated herein for the manufacture of a medication for eliciting an immune response in a mammal. In a particular (but non-limiting) embodiment, the medication so produced is a vaccine.

Additional non-limiting embodiments of the present disclosure are directed to a method of administering any of the pharmaceutical compositions disclosed or otherwise contemplated herein to a subject in need thereof. The amount of attenuated virus that is administered to a subject in need thereof varies according to many factors, e.g., the age, weight, overall health, gender, genetic history, history of allergies, prior infection, or vaccine history, etc., of the subject. The pharmaceutical compositions may be administered in a manner compatible with the dosage formulation and in such amounts as will be therapeutically effective (e.g., immunogenic and/or protective against infection with a wild type virus). The quantity to be administered depends on the subject to be treated, including, for example, the capacity of the immune system of the individual to synthesize antibodies, and, if needed, to produce a cell-mediated immune response. Precise amounts of active ingredient required to be administered depend on the judgment of the practitioner and may be monitored on a patient-by-patient basis. However, suitable dosage ranges are readily determinable by one skilled in the art and generally range from about 10^5 to about 10^6 plaque forming units (PFU) or more of virus per patient, more commonly, from about 10^5 to about 10^6 PFU of virus per patient. The dosage may also depend, without limitation, on the route of administration, the patient’s state of health and weight, and the nature of the formulation.

Upon inoculation with the pharmaceutical/vaccine compositions disclosed or otherwise contemplated herein, the immune system of the host can respond to the vaccine by producing antibodies, both secretory and serum, specific for the epitope(s) included in or expressed by the recombinant viruses. As a result of the vaccination, the host can become partially or completely immune to infection by the pathogen(s) carrying the epitope(s) or to wild type counterparts of the attenuated viruses that were injected. Where the epitope(s) is associated with human RSV (HRSV), the host may become resistant to developing RSV infection, or to developing moderate or severe RSV infection, particularly of the lower respiratory tract. The immune response may be innate or adaptive, and may be either cell-mediated or humoral. In a particular (but non-limiting) embodiment, the response is adaptive and leads to immunological memory. In a particular (but non-limiting) embodiment, the response is protective, i.e., the response prevents or at least lessens the impact of (e.g., avoids development of serious symptoms of) infection by other viruses with shared antigens and/or epitopes, e.g., other Pneumoviridae such as (but not limited to) wild type Pneumoviridae. Single or multiple administrations of the pharmaceutical composition disclosed or otherwise contemplated herein can be carried out. In neonates and infants, multiple administrations may be required to elicit sufficient levels of immunity. Administration can begin within the first month of life and continue at intervals throughout childhood, such as (but not limited to) at two months, six months, one year, and two years, as necessary to maintain sufficient levels of protection against the pathogen of interest. Similarly, adults who are particularly susceptible to repeated or serious infection by the pathogen of interest, such as (but not limited to) health care workers, day care workers, the elderly, individuals with compromised immune function, and individuals with compromised cardiopulmonary function, may require multiple immunizations to establish and/or maintain protective immune responses. Levels of induced immunity can be monitored by measuring amounts of neutralizing secretory and serum antibodies, and dosages adjusted and/or vaccinations repeated as necessary to maintain desired levels of protection.

Subjects who may be immunized using the formulations of pharmaceutical compositions disclosed or otherwise contemplated herein are usually mammals and are frequently humans, particularly human infants or children. However, this need not always be the case. Veterinary uses of the pharmaceutical compositions and methods disclosed or otherwise contemplated herein are also contemplated, e.g., for companion pets, or for animals that are of commercial value e.g., as a food source, or for any other animal, etc.

Further non-limiting embodiments of the present disclosure also include methods of eliciting an immune response to Pneumoviridae viruses in a subject or patient in need thereof. The method includes a step of administering any of the pharmaceutical compositions disclosed or otherwise contemplated herein to a subject. The method may include a step of identifying suitable recipients and/or of evaluating or monitoring the patient’s reaction or response to administration of the composition. In some embodiments, the composition comprises a live, recombinant attenuated mammalian (e.g., human) RSV (as described herein above or otherwise contemplated herein), and the subject is a child, an immunocompromised individual, an elderly patient, and/or any patient at risk of being exposed to RSV and developing an RSV infection. The method may be a method of vaccinating such individuals against developing severe (or alternatively, moderate) lower respiratory tract disease, e.g., against developing bronchiolitis.

The recombinant live, attenuated viruses disclosed or otherwise contemplated herein can also be used in diagnostic applications. In one non-limiting embodiment, a method useful for detecting the presence or absence of an antibody specifically reactive with an epitope is provided. The method includes the steps of contacting a sample with the recombinant virus carrying the epitope, and detecting any binding between an antibody component in the sample and the
recombinant virus. Examples of binding assays that are suitable for this purpose include (but are not limited to) ELISA (enzyme-linked immunosorbent assay), RIA (radio-immunoassay), FACS (fluorescence-activated cell sorter), and any combinations thereof.

Yet further non-limiting embodiments of the present disclosure include a method of generating antibodies specific for RSV in a mammal, wherein the method includes introducing into the mammal any of the recombinant, live attenuated viruses disclosed or otherwise contemplated herein (or any of the pharmaceutical compositions containing same, as disclosed or otherwise contemplated herein). Antibodies which specifically recognize one of the proteins or fragments thereof present in the virus may be used to detect production of the particular protein(s)/fragment(s), either in a laboratory setting (e.g., for research purposes) and/or to monitor infections established with the attenuated virus in a subject. Antibodies which specifically recognize the attenuated viruses disclosed or otherwise contemplated herein (both mono- and polyclonal) are also encompassed by the present disclosure. In some embodiments, antibody recognition is selective rather than specific. Antibodies may be polyclonal or monoclonal.

Certain additional non-limiting embodiments of the present disclosure are directed to a method of preventing or reducing the occurrence of respiratory syncytial virus infection in a mammal by administering any of the recombinant, live attenuated viruses disclosed or otherwise contemplated herein (or any of the pharmaceutical compositions containing same) to a mammal. In addition, any of the adjuvants described or otherwise contemplated herein may be administered simultaneously or partially or fully sequentially with the virus (or pharmaceutical composition containing same). In certain non-limiting embodiments, the mammal is susceptible to infection with RSV.

**EXAMPLES**

Examples are provided hereinafter. However, the present disclosure is to be understood to not be limited in its application to the specific experimentation, results, and laboratory procedures disclosed herein. Rather, the Examples are simply provided as one of various embodiments and are meant to be exemplary, not exhaustive.

**Example 1**

RSV is the most important viral respiratory pathogen of infancy and early childhood, and yet there is no approved vaccine. One of the main challenges thus far has been to achieve strong efficacy and safety within one vaccine. A trial in the 1960s using formalin-inactivated RSV vaccine (FI-RSV) failed to protect against RSC and even resulted in enhanced disease severity upon exposure to wild type RSV (termed vaccine-enhanced disease or VED). In RSV-naive children, VED is also induced by many subunit approaches but not by live-attenuated vaccines. Live vaccines have additional advantages over non-replicating or subunit vaccines in that they induce immunologically-balanced, longer lasting protection, including (but not limited to) mucosal immunity (natural site of infection) if administered intranasally.

To enable live vaccines with stable attenuation phenotypes, self-limited (single-round) live RSV viruses have been developed based on ablation of essential genes (Matrix [M] or Fusion [F] protein) by providing functional replacements via a complementing production cell line. As a demonstration, by pseudotyping RSV with baculovirus entry/exit protein GP64, it was possible to generate F-deleted yet infectious, single-round, RSV of high titer and with increased temperature stability (see, for example, US Patent Application Publication No. 2007/0104734 and U.S. Pat. No. 7,588,770, incorporated supra). Importantly, production of these GP64 pseudotypes is completely independent of F function. These viruses replicate their RNA genome at wildtype levels, generating abundant de novo viral antigens, but cannot spread beyond the initial site of infection. A similar experimental vaccine based on ablation of M induced robust immunity in an infant baboon model.

The spontaneous shift of the F protein from pre-fusion to post-fusion conformation during purification is believed to underlie the low levels of neutralizing anti-F antibodies induced by vaccine preparations, and probably also the loss of live RSV infectivity upon preparation and storage. Recent publications have shown that the F protein can be readily stabilized in the pre-fusion conformation through genetic changes, and when used as a protein vaccine, induced a higher proportion of neutralizing anti-F antibodies in vivo (see, for example, Kwong et al., incorporated supra). However, F is essential to RSV, and the genetically stabilized pre-fusion form (PreF) is no longer functional. A live vaccine expressing PreF in place of native F (to drive the immune response toward pre-fusion F without inducing VED in the RSV-naive population) is therefore not viable. Thus, to extend the advantageous pre-fusion F concept to the RSV-naive population, an F-independent production system is needed that allows generation of live RSV expressing PreF.

Thus, the present disclosure combines the pre-fusion F concept with the GP64 system previously developed by the inventor, as the GP64 system is F-independent and provides the tools to pursue these inventive concept(s).

**Example 2**

Oomens et al. (US Patent Application Publication No. 2007/0104734 and U.S. Pat. No. 7,588,770, incorporated supra) previously reported a baculovirus GP64 based complementation system that uniquely allows generation of infectious F-deleted or F-compromised viruses from cDNA in GP64-expressing cells. These GP64-pseudotyped viruses could be amplified to high titer and were significantly more temperature stable than wildtype RSV. Due to replacement of functional F with trans-complemented GP64, the viruses are infectious but self-limited and cannot spread beyond initially infected cells, thus constituting an attractive live-attenuated platform.

The present disclosure exploits this F-independent, GP64 complementation system to generate a live RSV which solely expresses a pre-fusion F protein variant. Replacing the native, functional F gene with a gene encoding a pre-fusion stabilized F in a live virus provides a novel combination of immunological benefits: it drives the anti-F response toward the pre-fusion F form, while also inducing a balanced response that includes cell-mediated immunity and avoidance of VED. In addition, a vaccine produced therefrom is single-round (self-limited) and thus cannot spread beyond the initially infected site, and is also more temperature-stable. Absence of the CT from preF also provides another safety advantage: if the live vaccine virus were to attempt to mutate preF in order to regain F function and virulence, absence of the CT will further prevent production of new progeny as the CT is required for virus assembly. The resulting immunogenic composition/vaccine thus has the
potential to exceed previous formulations in inducing a broadly efficacious yet safe immune response for the RSV-native target population.

In certain non-limiting embodiments, the present disclosure uses the F-independent system based on a cell line that provides baculovirus GP64 in trans to RSV, to generate live viruses that express a non-functional pre-fusion F protein variant. In this manner, not only will the humoral arm be activated, leading to anti-pre-fusion F antibodies, but the cellular arm will also be activated, leading to anti-pre-fusion F CD8\(^{+}\) lymphocytes, among others. Thus, contrary to protein-based vaccines, the live immunogenic compositions/vaccines of the present disclosure will elicit both a humoral and a cellular response. Because the immunogenic composition/vaccine is based on RSV itself, immunity will also be induced against all other RSV antigens, including (but not limited to) G. The F-independent system provides the baculovirus GP64 protein in trans, which results in a virus that is infectious but only for a single-round, thereby making the virus incapable of inadvertent spreading through the lung of a recipient, and thus safer.

Three versions of pre-fusion stabilized F protein variants were generated for use in accordance with the present disclosure, as shown in FIG. 1. These pre-fusion stabilized F protein variants are based on the previously described pref fusion protein variant DS-Cav-1 (see, for example, US 2015/0030622, US 2016/0031972, and US 2016/0046675, incorporated supra; and McLellan et al. (Science (2013) 342:592-598); the entire contents of which are expressly incorporated herein by reference). Pref\(^{SCFCT}\) is a membrane-anchored version that is expressed and anchored at the surface of infected cells; the amino acid sequence of pref\(^{SCFCT}\) is represented by SEQ ID NO:2. Pref\(^{SEC}\) is a secreted version that is secreted to the extracellular environment on infected cells; the amino acid sequence of pref\(^{SEC}\) is represented by SEQ ID NO:3. Pref\(^{SEC\text{cyt}}\) is similar to Pref\(^{SEC}\) but contains an epitope tag for easy identification and detection; the amino acid sequence of pref\(^{SEC\text{cyt}}\) is represented by SEQ ID NO:4.

RSV viruses were then engineered with one of the three pre-fusion stabilized F variants of FIG. 1 inserted at either the 8\(^{th}\) or 6\(^{th}\) genome position, as shown in FIG. 2. Panel B of FIG. 2 depicts RSV genomes with variants of pre-fusion stabilized F at the 8\(^{th}\) genome position, while Panel C of FIG. 2 depicts RSV genomes with variants of pre-fusion stabilized F at the 6\(^{th}\) genome position. The 6\(^{th}\) genome position was more highly expressed than the 8\(^{th}\) genome position, to enhance the level of pre-fusion F. In addition, all of these viruses also contained a GFP (Green Fluorescent Protein) marker gene for tracking and assay purposes. However, it will be understood that the presence of GFP was simply for experimental purposes; GFP is not required to be present in the viruses of the present disclosure and can be removed if necessary. All of these viruses also lacked expression of the secreted G protein (indicated as Gnem), which is a known virulence factor. These vaccine viruses have been successfully generated, and stocks for each have also been generated with titers over 10\(^7\) plaque-forming units per ml.

Successful expression of pre-fusion stabilized F protein variants from the engineered viruses generated as in FIG. 2 was verified, as shown in FIG. 3. Cells infected by virus RSV8-pref\(^{SCFCT}\) were incubated with anti-F and anti-G antibodies at 26 hours post-infection, or mock-infected as a negative control, and subjected to ELISA. Three antibodies (provided by J S McLellan, The University of Texas at Austin) were used to detect the presence of F protein. The first antibody, Motavizumab (mota), detects both the pre-fusion and post-fusion conformation of F, while the second and third antibodies, D25 and 14402, are known to detect a different epitope specific only for the pre-fusion conformation of F (site \(\Phi\) and site V). In addition, the G protein was detected at similar levels.

Therefore, as can be seen in FIGS. 4 and 5, abundant levels of pre-fusion F were expressed at the surface of virus-virus infected cells. In addition, Viruses RSV8-pref\(^{SEC}\) and RSV8-pref\(^{SEC\text{cyt}}\) have been similarly examined and were also demonstrated to express pre-fusion F.

In addition, viruses with pre-fusion F variants at position 6 will generate higher levels of pre-fusion F.

It was noted that pre-fusion F protein variants that contained the cytoplasmic tail of the F protein do not express at the cell surface as well as the wildtype F protein. However, when the cytoplasmic tail was removed (such as (but not limited to) in virus RSV-pref\(^{SCFCT}\) described in FIGS. 1-3), surface expression was improved to an expression level similar to that observed for wildtype F protein. Therefore, while the scope of the present disclosure includes the use of F protein variants both with and without cytoplasmic tails, the absence of the cytoplasmic tail can improve surface expression of the pre-fusion F protein variant and can thus assist with inducing immunity against the pre-fusion F conformation.

Example 3

Removal of the cytoplasmic tail (CT) strongly improved cell-surface expression of pre-fusion F. HEP2 cells were transfected with the indicated F expressing plasmids. The F open reading frames were codon-optimized, as native F sequences express poorly in transfected cells. To each well, a plasmid expressing NGFR-myc was added as a transfection control (NGFR-myc is expressed at the cell surface). At 46 hours post-transfection, transfected cells were incubated with various F antibodies or myc antibody as a control, and relative F surface levels were detected using standard ELISA.

As shown in FIG. 3, full-length F and pref\(^{F}\) were detected at the cell surface equally and at low levels. However, removal of the cytoplasmic tail (CT) from pref\(^{F}\) (pref\(^{SCFCT}\)) led to a strong increase in surface expression. As expected, surface expressed pref\(^{F}\) was recognized by pre-fusion-specific site 0 and V antibodies but not by a post-F specific antibody, demonstrating that pref\(^{SCFCT}\) is in the pre-fusion conformation.

The vaccine candidate RSV6-pref\(^{SCFCT}\) induced high surface levels of prefusion-F and G. HEP2 cells were infected with the indicated viruses. At 26 hours post-infection, infected cells were incubated with F, G, or N antibodies, and relative F and G surface levels were determined using ELISA (the N protein is an indicator of viral genomic replication and is shown for normalization purposes; to detect N, cells are detergent-permeabilized).

As can be seen in FIG. 5, RSV6-pref\(^{SCFCT}\) induced higher surface expression of prefusion F and G than a recombinant wild type (WT) virus. Pref\(^{F}\) is recognized by pre-fusion-specific site 0 and V antibodies but not by a post-F specific antibody, demonstrating that pref\(^{SCFCT}\) is in the pre-fusion conformation. As expected, RSV6-pref\(^{SEC}\) expresses G but not pref\(^{F}\) at the plasma membrane.

In addition, vaccine candidate RSV6-pref\(^{SEC\text{cyt}}\) secreted high levels of prefusion F. HEP2 cells were infected with the indicated viruses. At 36 hours post-infection, supernatants of infected cells were harvested and incubated on ELISA plates.
coated with the anti-tag antibodies for 1 hour. Bound preF* was then detected by ELISA using D25 and motavizumab as primary antibodies.

As shown in FIG. 6, RSVV-pref*ΔNClog secreted high levels of fusion F, which was recognized by pref*-specific antibody D25. As expected, RSVV-pref*ΔNC did not secrete any pref fusion F protein into the supernatant. Strong but slightly lower levels of pref F were detected at 24 hours post-infection.

Pref* expressing single cycle RSV was also shown to induce high levels of anti-RSV antibodies in vivo. 96 well plates were coated with pref*G+4 by infecting HeP-2 cells with RSVV-pref*ΔNC. At 26 hours post-infection, pref* and G proteins were present at the cell surface in conformationally accurate (native) form. Pooled sera (n=3, collected at 3 weeks post-boost) from mice vaccinated prime/boost with RSVV-pref*ΔNC or Rec WT virus were incubated on the coated ELISA plates, and antibody levels were determined using ELISA.

As shown in FIG. 7, prime/boost vaccination with RSVV-pref*ΔNC induced anti-pref antibody levels similar to a wildtype virus, despite being limited to a single cycle of replication. (Note: The shown pref* vaccine was codon-optimized, codon-optimized and non-codon-optimized pref* have been tested and gave similar results in mice.) A low dose (0.5 million PFU/vaccination) of RSVV-pref*ΔNC induced equal levels of antibodies as a high dose (1 million PFU/vaccination).

Example 4

In this Example, different pref*-based single cycle RSV vaccines were constructed, and FIG. 8 graphically depicts a schematic overview of five examples of different pref*-based single cycle RSV vaccines constructed in accordance with the present disclosure. In addition to RSVV-pref*ΔNC, two types of pref* based vaccine candidates were generated. First, vaccines were generated in which known viral virulence factors NS1 or NS2 have been moved to downstream positions to downregulate their expression levels. NS1 and NS2 are known to block the host interferon response, and downregulating their expression is expected to alter and improve the quality and longevity of the immune response. As such, NS1 or NS2 were separately moved to the 5th genome position. Second, vaccines were generated in which pref*ΔNC was moved to the 1st or 2nd genome position, for enhanced expression. The specific vaccine candidates generated are shown in FIG. 8 and include: First panel, RSVV-pref*ΔNC; second panel, RSVV-pref*ΔNC,NS1low; third panel, RSVV-pref*ΔNC,NS1low; fourth panel, RSVV-pref*ΔNC,NS1low; fifth panel, RSVV-pref*ΔNC,NS2low. RSVV-pref*ΔNC and RSVV-pref*ΔNC,NS1low have been examined in vitro and in vivo, as shown in FIGS. 9-12 and described in detail herein below.

FIG. 9 graphically depicts that different pref* RSV vaccines induced high levels of pref and G protein at the cell surface. HeP-2 cells were infected with viruses RSVV-pref*ΔNC and RSVV-pref*ΔNC,NS1low. At 26 hours post-infection, infected cells were incubated with anti-pref and anti-G antibodies, which were subsequently detected by standard ELISA method. Anti-N antibody was also used as an indicator for viral genomic replication (N encapsulates the viral genome), and pref and G antibody levels were determined without and with N level-based normalization.

Judged by N level, virus RSVV-pref*ΔNC,NS1low replicated to lower levels than RSVV-pref*ΔNC. This matches the observation that RSVV-pref*ΔNC,NS1low spreads a little more slowly through a cell culture, as seen by GFP expression. This is also consistent with the literature and with NS1 having both anti-immune and pro-viral functions. Judged by normalized N levels, on a per virus basis, the two viruses generated very similar levels of pref and G proteins. This indicates that moving pref*ΔNC to the first genome position did not raise pref levels, counter to expectations.

Next, the two viruses were examined in vivo. As can be seen in FIG. 10, different pref RSV vaccines induced high levels of pref-specific and G-specific antibodies. 96 well plates were coated with pref*G or pref* alone as follows: HeP-2 cells were infected with RSVV-pref*ΔNC or RSVV-pref*ΔNC,AG (G gene removed). At 26 hours post-infection, either pref*G or pref* alone were present at the cell surface in conformationally accurate (native) form. Pooled sera (n=3, collected at 3 weeks post-boost) from mice vaccinated prime/boost with RSVV-pref*ΔNC and RSVV-pref*ΔNC,NS1low were incubated on the coated ELISA plates (pref* alone on the left; pref* alone on the right), and antibody levels were determined using standard ELISA method. Anti-G Ab L9 was used to verify the absence of G protein in the pref*-alone ELISA.

Prime/boost vaccination using two distinct single cycle pref* vaccines (RSVV-pref*ΔNC and RSVV-pref*ΔNC,NS1low) both induced anti-G and anti-pref* antibodies, despite being limited to a single cycle of replication. Whereas RSVV-pref*ΔNC,NS1low expressed in cell culture overall lower pref and G levels than RSVV-pref*ΔNC, it induced equal levels of pref* antibodies in vivo and moderately high levels of anti-G antibodies than RSVV-pref*ΔNC, indicative of potential vaccine advantages. Lower NS1 levels may also increase immune memory.

The two distinct pref* RSV vaccines were also shown to induce higher neutralizing antibody activity than a wildtype virus, despite being safe, single-cycle vaccines. Neutralizing anti-F antibodies from mice vaccinated with RSVV-pref*ΔNC, RSVV-pref*ΔNC,NS1low, or rec WT (ELISA) were tested. Three-fold dilutions of pooled mice sera (3 mice per pool; sera harvested 3 weeks post-boost) were incubated with 500 PFU (FIG. 11) or 250 PFU (FIG. 12) of virus RSV-AG-HRP, which lacks the G protein (allowing detection of F-specific neutralization) and contains the HRP gene for detection. After a one hour incubation, virus-antibody suspensions were incubated on HeP-2 cells for 1.5 hours. Inoculum was removed and cells incubated for a total of 48 hours post-infection (hpi). At 48 hpi, medium was replaced with standard ELISA substrate, and OD450 was determined after 30 minutes as a measure of virus replication.

As shown in FIGS. 11 and 12, all viruses induced F-specific virus-neutralizing antibodies. RSVV-pref*ΔNC and RSVV-pref*ΔNC,NS1low induced slightly higher levels of F-specific neutralization than rec WT virus, despite being limited to a single cycle of virus replication. Neutralizing serum antibodies are a strong predictor of in vivo protection potential. Because rec WT virus was previously shown to protect mice from RSV challenge, the novel vaccines of the present disclosure will protect animals in vivo and will also be safe, since they only replicate for a single round.

Thus, in accordance with the present disclosure, there have been provided compositions, as well as methods of producing and using same, which fully satisfy the objectives and advantages set forth hereinabove. Although the present disclosure has been described in conjunction with the specific drawings, experimentation, results, and language set forth hereinabove, it is evident that many alternatives, modifications, and variations will be apparent to those skilled in the art. Accordingly, it is intended to embrace all such alternatives, modifications, and variations that fall within the spirit and broad scope of the present disclosure.
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| Thr Val Ser Val Gly Asn Thr Leu Tyr Val Tyr Val Asn Lys Leu Glu Gly | 450 | 455 | 460 |
| Lys Asn Leu Tyr Val Lys Gly Glu Pro Ile Ile Asn Tyr Tyr Asp Pro | 465 | 470 | 475 | 480 |
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| Tyr Gly Lys Thr Lys Cys Thr Ala Ser Asn Lys Asn Arg Gly Ile Ile | 420 | 425 | 430 |
| Asp Ile Ser Ser Ser Val Ile Thr Ser Leu Gly Ala Ile Val Ser Cys | 405 | 410 | 415 |
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| &lt;213&gt; ORGANISM: Artificial Sequence |   |   |   |
| &lt;220&gt; FEATURE: OTHER INFORMATION: Variant of Respiratory Syncytial Virus (RSV) F protein |   |   |   |
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| Ile Asn Ala Leu Tyr Leu Thr Ser Ser Glu Asn Ile Thr Glu Glu Phe | 20 | 25 | 30 |
| Tyr Gln Ser Thr Cys Ser Ala Val Ser Arg Gly Tyr Leu Ser Ala Leu | 35 | 40 | 45 |
| Arg Thr Gly Trp Tyr Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile | 50 | 55 | 60 |
| Lys Glu Thr Lys Cys Asn Gly Thr Asp Thr Lys Val Lys Leu Ile Lys | 65 | 70 | 75 | 80 |
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| Ile Ser Lys Lys Arg Lys Arg Arg Phe Leu Gly Phe Leu Leu Gly Val | 130 | 135 | 140 |
| Gly Ser Ala Ile Ala Ser Gly Ile Ala Val Cys Val Leu His Leu | 145 | 150 | 155 | 160 |
| Glu Gly Glu Val Asn Lys Ile Lys Asn Ala Leu Gln Leu Thr Asn Lys | 165 | 170 | 175 |
| Ala Val Val Ser Leu Ser Asn Gly Val Ser Val Leu Thr Phe Lys Val | 180 | 185 | 190 |
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cgcgcgcccg gcacacgctgc gcgcctgctg tntatacgac gcgcctgtttc gcgcctgtttc 600
aaccagacgg gcacacgttgc gcgcctgtgc tntatacgac gcgcctggtc gcgcctgtttc 660
atcgagttcc agcagaagaa caaocgcctg ctggagactca ccocgcagtt cagctgtaac 720
gcggcgtgta caaccccccgt gaccaactac atctgaccca caacgcagct cgtgacccgt 780
atcagacga taccccatca caagcagcag aagaagcgtga tgcagcagaca caccgagtc ac 840
gtgcgcgaag cacgctaggc ccatcgtgt atccacgaag aggagttgtg cgcctgctgt 900
gtgcagctgc cccgtgctag ccggcagcag aacccctgtg ggaagctgca caccagcccc 960
tctggcaccaca cccaccccaac gaggcgcgcg acactctgcac tgcagtctac cgcagccggc 1020
tgttactcgcc acaagcgcgga cagcgtgacattctccccca aagcgcagac ctggaaagtg 1080
cragaccaac ggctgtgcttg cagacacattg aacagcgcgta cctgtgcacag cgaagtgcaac 1140
cctggcacaag tcgacatctt caaocccaaat gaactacgca aagcatcagc caccgaagcc 1200
gacgtgcacga ggcgctgctg caccagcctgg ggcgtccagtc tgacgtcaat ggcgaagacc 1260
aagtgcacgg gacgacacac aacgacg aggctgacaca atcatacagg ccttcgacgaa eggtgaccc 1320
ctcgtgtaac acaagcgcgct ggcacgctgt agcgggctga aacaccgttgta ctcagtgac 1380
aagcagaggg ggcagtgacct gtagtgccag ggagagccgca tcataccatt tctagccca 1440
cctgggtcttc caaagcggaaaa tggtagacg cagcagcagt aagctgaaaga gaagccatac 1500
cagagctctgg cttccctcagc caagacagcc gcgcgtgctgg gccgag 1545

<210> SEQ ID NO 7
<211> LENGTH: 1572
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Codon-optimized sequence encoding RSV F protein variant of SEQ ID NO:4

<400> SEQUENCE: 7

atggagcctgc tgaatcctgg gcgcaacgcgc atcacaacata ctcgtgaccgc ctagcacttc 60
tgctggcaca ggcgctcagaa cacaaccagag ggtctctaac aagcagcctgt cagcgcctgtg 120
agcagggccagct acgcgcgact cctgcgacacc ggcctggaac caaagctgtag cacaatagcag 180
tcgcgacacaa ccaaggtgaa aacagtgaaca gcacgcagag ccaaaagtgg gtagtcaag 240
cacaagctgg acagaatcaca gcagcgcgttg agcagctgtg acgcgctgac ccacagtcac 300
ccgcgcacca acaacggcgcgc cccgcgctcg ctggcagctc tctgatcact caccctgcag 360
aagcacaaga agaagccagtt gcacgtgacg aagacgagca agcgcgtcgt ctcggctct 420
tcctggtggtcg tgtgctcaggg cactgcgagg cggctgtggcc tggctaatgt gctgcccttg 480
ggaggggaaag tgaacagcgcgc ctgtgcgacac ccacaacagggctcg cttgatgacg 540
ttcgcgacag cttgtggtcgt gctgtccttc aacagtgcgtg acagtgaaag ctacattgcac 600
aagctgcctgc tgcagcatctc cacaacagcag aagctgagcgc ctaaagcact ctgagcctgt 660
atcgagttcc agcagaagaa caaocgcctg ctggagactca ccocgcagtt cagctgtaac 720
gcggcgtgta caaccccccgt gaccaactac atctgaccca caacgcagct cgtgacccgt 780
atcagacga taccccatca caagcagcag aagaagcgtga tgcagcagaca caccgagtc ac 840
gtgcgcgaag cacgctaggc ccatcgtgt atccacgaag aggagttgtg cgcctgctgt 900
gtgcagctgc cccgtgctag ccggcagcag aacccctgtg ggaagctgca caccagcccc 960
tctggcaccaca cccaccccaac gaggcgcgcg acactctgcac tgcagtctac cgcagccggc 1020
tgttactcgcc acaagcgcgga cagcgtgacattctccccca aagcgcagac ctggaaagtg 1080
cragaccaac ggctgtgcttg cagacacattg aacagcgcgta cctgtgcacag cgaagtgcaac 1140
ctgctgcaag tggacctctt caaccccaaag taagacgtac aatcatacag cagcaagacc 1200
gagctgcaag gcagcctgctt cacagctctg ggcgccattg tcaagctgta caaggggagc 1260
aagttgacac ccagacacaa gaacgagcggcg acatcaagctt ccctcagcaaa cggctgcgac 1320
tacgctgacca acaagggcctt ggacacccgtg aagctgggaaga acaaccctgta ctacgtgaac 1380
aagcagaggg ggaagagctt ctaagttgac ggagcagcct ttcaatactct tcaagacccc 1440
tctgctgctc cccgcagcag tttgtacgac ccagacccgc aagttgacag gaagatcaac 1500
cagagtcttg cctctacgcc caagagcctg cagctgcttg ggcagctggaa ggaagccgac 1560
ggctggagct ag 1572

<210> SEQ ID NO: 8
<211> LENGTH: 1665
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Gene encoding RSV F protein variant of SEQ ID NO:2, based on wild type RSV nucleotide sequence

<400> SEQUENCE: 8

atggagtgcc taacctctca aagcaatgca attaccccaa ttcctacttcag agtcacatctt 60
tgtttttgctt ctcgctcaaaaa ctaactgtaaa gaatatttct aatcatacagc cagctgcagtt 120
agcacaaggt atcttactcg tctggaacct ggggttgtata ccagtggtat aacctatgaag 180
ttaagtaata tcaagaaaat ctcatgagtt gaacagatgt ctcaagtttaa attgataaaa 240
cacaagatag ataatatatataa aatactgtc acagtgttcagc caggaagcaac 300
cacccaaacc aacagctgcacc gagaagagag cagcaagctct ctagctctcct 360
aagcttccaa aaactttcaag aagctggagc aagaaagaata aagagatctttctctgatt 420
tggtaggtgct tggtagtctgct atagccagatgg ggcgttagtgct atagccagtt ctgcaacta 480
gagggggaag ggaagagactt ccaaaatgt ccatactcaca ccagagagcctcgagtcacg 540
tttcatataag gtagttgctt ottaacctctt aaattgcttag acctcaaaaa ctatataagat 600
aaacaaaatgt atccatattct ccagccaacaa ggtcagccgta tataaatatatgag 660
ataaggcttc acaaaagga ccaagacttc ataggtgattt ccaggggaattt taggttttattt 720
gaggtggat tcatactcgtaa aatcagattc atgtcaatgtagaat tttacctagatt 780
ataacgtaata tggagaatag ctacatctcctt cagttgactagttaaag ttagcaacaca 840
gttgagagc aagttactttc tattgctgtc atataaaaag aggagctctt tcatatagta 900
gtcaatcttc cctttctgtct gcagattagctt gggctggttaa cccagagctcc 960
cctctctgtc cacacatag cgcagcagat tctcagtcgcata aatcagttgaa cccagagctcc 1020
tgaggctgc gctgtgaatgtc aatcagttgaa cccagagctcc 1080
cacccaaacc aagecagatgc cagctgatca caagcatagtctt cagctgatca 1140
cctcgcacttg tgcagtaattt ccaccccaaaaa tttctgcttat aacatgcttgctcctgc 1200
gagctgcaag gcagcctgctt cacagctctg ggcgccattg tcaagctgta caaggggagc 1260
aagttgacac ccagacacaa gaacgagcggcg acatcaagctt ccctcagcaaa cggctgcgac 1320
tacgctgacca acaagggcctt ggacacccgtg aagctgggaaga acaaccctgta ctacgtgaac 1380
aagcagaggg ggaagagctt ctaagttgac ggagcagcct ttcaatactct tcaagacccc 1440
tctgctgctc cccgcagcag tttgtacgac ccagacccgc aagttgacag gaagatcaac 1500
cagagtcttg cctctacgcc caagagcctg cagctgcttg ggcagctggaa ggaagccgac 1560
ggctggagct ag 1572
agcaacacca acactatatg cacacaccac tctacttgta tctaggtgat ctctgtgac 1620
cctgctgctg tggctgtctgt gctgtactgc aagcaccgca gaatg 1665

<210> SEQ ID NO 9
<211> LENGTH: 1545
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Gene encoding RSV F protein variant of SEQ ID NO:3, based on wild type RSV nucleotide sequence

<400> SEQUENCE: 9
atggagttgc taatcctcga agcaatgca attaccacaa tcctcactgc agtacacattt 60
tgttttgctt ctgtgctaaaa ctcacagtaaa gaatttttac aatcaacagc cagttgcatgt 120
agcaaaagtct atctagtgc tcctgagaact gttggtttaca acctctgtat acatcgataa 180
ctacactata tcacagtaaa taatgttata ggacacgtat ctaagtttaaa attgataaaa 240
cacaagttg ataatatat aa tactgttgaa acaagagtcg agtgcctcac gcgaagcaca 300
ccagcaacca acactctgact caggaagggaa tcacacagctg tgaagttttt 360
aatgcaccaaa aacacacttg acattaagct gccaaagggag aagaagattt ctctgtggtttt 420
tctgtgtgctt ttgagtctgc attcgccagt ggccctgctgt tagcaggtt cctgacactctta 480
gaggggagacgccaagagct ccaagtacct ctacactaca caacacaggc ttagttcgcg 540
ttatcaacat ggttagttg cttttaccttc aataagttgtag accctaaaaa cttatagat 600
aaacatggtct atctactttc ccaagaccct aacaaatagt taaatagcttg 660
atagagcctc aacacagatat cagacactta ctagagtata cccggaatct tagttgtaaat 720
gctgatcc ttcagccagg ttagaactac atgttcaatc cctgtttaaatgctt 780
atctagata tgcctataac aaacttgcat cccaaaattgctttgc cttctgaaataa 840
gttsagacag aaggttcttt aacagtctgat taattaaaccta agaagactttt ccatcattga 900
tgacactcct ctttattagtg ttgtagatag accctctgctt ggacacatca cca ctaccccctc 960
ctacttagct caacacacaa aagaaaaaac aacatgcttt taacaagactg tggagggga 1020
tggtctgtg acacacgacg aactgcagtt ttctctccac agctggaacc atgtaaagtt 1090
catcaacatc ggttgctttg tgcacaacaag cagttcttta cattacagct tggaggaat 1140
tctgtgatct cactgacct acaaaaaac aagttcagaac attcgctcag tggagggaga 1200
gattagccca gcctctttctt ccacactcact gcggcatctt gctctagcaag tggaaaaattct 1260
aatgtaacg ccaccataaa aacatgctaga atctataagc aatattctaaag cgggttgcag 1320
ctctgatcc aatgaggggt gcagactgtt tcctgtgttag acaactattg tttgcttaaat 1380
aaggagagag agttggcattt ccagctagttt cttcagcaatc tttagaaattt ctatgtgcc 1440
ctactgctt ccctcctagta aatggtgca ccaatcactc aacaaagcgaag aagattttc 1500
cyctggtgctt ccactattcgc cagaggcagc ggcctgtctgt gtata 1545

<210> SEQ ID NO 10
<211> LENGTH: 1572
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Gene encoding RSV F protein variant of SEQ ID NO:4, based on wild type RSV nucleotide sequence

<400> SEQUENCE: 10
atggagttgc taatcctcga agcaatgca attaccacaa tcctcactgc agtacacattt 60
\texttt{tgtttgcct ctggctaaaa cactacgaa gatttttact aactcaacag cagtcgagtt}
\texttt{120}
\texttt{agcaaaaggtct acctttgctc tgtgagatct ggtttgatata ccagcgttat aactatagaa}
\texttt{180}
\texttt{tttaagataa tcaagaaaaa taagtgtaaat gaaacagatgc taagttgacaa attgataaaa}
\texttt{240}
\texttt{caaaatattag ataaataataa aaagttgtga acagaaattgac agttgtccac gcagaagcaca}
\texttt{300}
\texttt{ccagcacaac acaatcagac cagaagagaa atcaacaaagtt cctatgaatt tacaactcaac}
\texttt{360}
\texttt{aattgcaaaa aaccaagatt acataatagc aagaaaaaga aagaaagatt ctcttggtt}
\texttt{420}
\texttt{tctgagttgct tgtgagatcgc aatcggcagtt ggctttgcttg tgtgaagagtt ctctgcaacct}
\texttt{480}
\texttt{gaagggggaa ctgacaagat caaaggtgct ctactacccca cacaagagcg tctagtca}
\texttt{540}
\texttt{tttatcaagt gatgttagctg tctaaacctc aacggtttag acctcctaaa atatatagat}
\texttt{600}
\texttt{aaacaaatgt taccctaatt caacaagcaca agtggcaca taataanat agaaactgtg}
\texttt{660}
\texttt{ataagattcgc aacaagaaaga cacaagacta tctagagatta cccggaatatt taggtttaat}
\texttt{720}
\texttt{gcaggttgaat tcaacagctt gtagaatcct acgtctaaactt atagtggaatt atgtcatta}
\texttt{780}
\texttt{attactgcttc tggcttatataa aataagctac gaaagtttacattatcgac cacaatccatt}
\texttt{840}
\texttt{gtttgaagct aagttatcctt atcatgctt atataaagaa aagaggttatt atcagatgt}
\texttt{900}
\texttt{gttaacacttc ctggtaatag tgcgctcaac atacgctgtt ggaactaca cacaactcct}
\texttt{960}
\texttt{ctattgacga acaaaccaaaa agaaggttcc aacctcgttt taacaagacac tgcagagaga}
\texttt{1020}
\texttt{tggtactagt gcagacgagg tctgtactct tccctccacac aagtgtaaacag atcaagttt}
\texttt{1080}
\texttt{caatatcagc gatgtatttg tgcacaagt aataagttta cattccaaag tgaagtaaat}
\texttt{1140}
\texttt{ctcgcgatg tgtcaratat caacocccaaa tctgattgta aaatttgcac ttcacaaaaa}
\texttt{1200}
\texttt{gatggaagct ctcgcgatg tccatcttc aagtttttgg tctagttgta tgggaaactt}
\texttt{1260}
\texttt{aaatgtcagc catcctaaac aatactgtgga atccatagaa ctttcttatt ccggttgctg}
\texttt{1320}
\texttt{tctatctaa ataagaaggttg gcagctggct tgtctaggtta acacattata ttatgtaaat}
\texttt{1380}
\texttt{aagcaagaggtttcagctt ctgatgatagc atggataaaggt cctagcttgagcaga cacaaga}
\texttt{1440}
\texttt{ttgatctctt ccttcaagta atttggagca tcaatatctt aagcttacga gaagataaac}
\texttt{1500}
\texttt{ccagagtattgc ctccttcagcg ccagagccag gacgctgtgg gcagcttgaa gaagagccggcag}
\texttt{1560}
\texttt{gggctggact aa}
\texttt{1572}

\texttt{<210> SEQ ID NO 11}
\texttt{<211> LENGTH: 9}
\texttt{<212> TYPE: PRT}
\texttt{<213> ORGANISM: Autographa californiae nucleopolyhedrovirus}

\texttt{<400> SEQUENCE: 11}

\texttt{Ser Trp Lys Amp Ala Ser Gly Trp Ser}
\texttt{1 5}

\texttt{<210> SEQ ID NO 12}
\texttt{<211> LENGTH: 27}
\texttt{<212> TYPE: DNA}
\texttt{<213> ORGANISM: Autographa californiae nucleopolyhedrovirus}

\texttt{<400> SEQUENCE: 12}

\texttt{agttggaag agccagcgg gttgagc}
\texttt{27}

\texttt{<210> SEQ ID NO 13}
\texttt{<211> LENGTH: 298}
\texttt{<212> TYPE: PRT}
\texttt{<213> ORGANISM: respiratory syncytial virus}
<210> SEQ ID NO 15
<211> LENGTH: 520
<212> TYPE: PRT
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: Chimera of Baculovirus GP64 protein with cytoplasmic tail of RSV F protein

<400> SEQUENCE: 15

Met Val Ser Ala Ile Val Leu Leu Ala Ala Ala Ala His  
1        5       10       15
Ser Ala Phe Ala Ala Glu His Cys Asn Ala Glu Met Lys Thr Gly Pro  
20       25       30
Tyr Lys Ile Lys Asn Leu Asp Ile Thr Pro Pro Lys Thr Leu Glu  
35       40       45
Lys Asp Val Glu Ile Thr Ile Val Glu Thr Asp Tyr Aen Glu Aen Val  
50       55       60
Ile Ile Gly Tyr Lys Gly Tyr Tyr Glu Ala Tyr Ala Aen Gly Aen Gly  
65       70       75       80
Ser Leu Asp Pro Asn Thr Arg Val Glu Glu Thr Met Lys Thr Leu Aen  
85       90       95
Val Gly Lys Glu Asp Leu Met Trp Ser Ile Arg Glu Glu Gly Cys Glu  
100      105      110
Val Gly Glu Leu Ile Asp Arg Trp Gly Ser Asp Ser Asp Asp Cys  
115      120      125
Phe Arg Asp Aen Glu Gly Arg Gly Glu Trp Val Lys Gly Lys Glu Leu  
130      135      140
Val Lys Arg Glu Aen Aen Aen His Phe Ala His His Thr Cys Aen Lys  
145      150      155      160
Ser Trp Arg Cys Gly Ile Ser Thr Ser Lys Met Tyr Ser Arg Leu Glu  
165      170      175
Cys Glu Asp Aen Thr Asp Glu Cys Glu Val Tyr Ile Leu Asp Aen Glu  
180      185      190
Gly Asn Pro Ile Aen Val Thr Val Aen Thr Val Leu His Arg Asp Gly  
195      200      205
Val Ser Met Ile Leu Lys Gly Ser Thr Phe Thr Thr Arg Glu Ile  
210      215      220
Lys Ala Ala Cys Leu Leu Ile Lys Asp Aep Lys Aen Aen Pro Glu Ser  
225      230      235      240
Val Thr Arg Glu His Cys Leu Ile Asp Aen Asp Ile Tyr Asp Leu Ser  
245      250      255
Lys Aen Thr Trp Aen Cys Lys Phe Aen Arg Cys Ile Lys Arg Lys Val
Glu His Arg Val Lys Lys Arg Pro Pro Thr Trp Arg His Asn Val Arg
260       280       285
Ala Lys Tyr Thr Glu Gly Asp Thr Ala Thr Lys Gly Asp Leu Met His
275       290       295
Ile Gln Glu Glu Leu Met Tyr Glu Asn Asp Leu Leu Lys Met Asn Ile
305       310       315
Glu Leu Met His Ala His Ile Asn Lys Leu Asn Asn Met Leu His Asp
325       330       335
Leu Ile Val Ser Val Ala Lys Val Asp Glu Arg Leu Ile Gly Asn Leu
340       345       350
Met Asn Ser Ser Val Ser Ser Thr Phe Leu Ser Asp Asp Thr Phe Leu
355       360       365
Leu Met Pro Cys Thr Asn Pro Pro Ala His Thr Ser Asn Cys Tyr Asn
370       375       380
Asn Ser Ile Tyr Lys Glu Gly Arg Trp Val Ala Asn Thr Asp Ser Ser
385       390       395
Gln Cys Ile Asp Phe Ser Asn Tyr Lys Glu Leu Ala Ile Asp Asp Asp
405       410       415
Val Glu Phe Trp Ile Pro Thr Ile Gly Asn Thr Thr Tyr His Asp Ser
420       425       430
Trp Lys Asp Ala Ser Gly Trp Ser Phe Ile Ala Gln Gln Lys Ser Asn
435       440       445
Leu Ile Thr Thr Met Glu Asn Thr Lys Gly Val Gly Thr Ser
450       455       460
Leu Ser Asp Ile Thr Ser Met Ala Glu Gly Leu Ala Ala Lys Leu
465       470       475
Thr Ser Phe Met Phe Gly His Val Val Phe Val Ile Leu Ile
480       490       495
Val Ile Leu Phe Leu Tyr Cys Met Ile Ser Arg Arg Gln Leu Ser Gly
500       505       510
Ile Asn Asn Ile Ala Phe Ser Asn
515       520

<210> SEQ ID NO 16
<211> LENGTH: 1563
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE: OTHER INFORMATION: Nucleotide sequence encoding the amino acid sequence of SEQ ID NO:15
<400> SEQUENCE: 16
atggtaaagcc caatgtgtttt atatggtcatt ttggcggcgg gggcgcattt tgcatttgccg
60
gcggagact gcacgcgca aataagagc gtcgctaca agattaaaaa cttgacatt
120
accccgccgca aggsaagtct gcacacagac gttgaatcta ccctcgttgta gcagcactac
180
aacgcsaacc acgatattcg gctcaggggg tactaccagg cgatgctga caacggcggc
240
tgcgctgac ccacacacag cgtcgaagaa accatgaaaa cgtgagtgtg gggcgaagag
300
gacttctca tggctgagct cacagcagcg tggaggttgg gcagagacgt gatgagacgt
360
tgggagcttg acaggcgcag cgtgttttgc gacaagagt ccggcgcag gtcgctgtagc
gcagagcgag aatccttttg cgcacacac gtcgactaa
540
acggacaggt gcacagttata catattggac gcctgagggca aaccccaataa cctgcaccctg
600
gacaatctgcg ttcatatgca cgggtctgagc atgatttcaa aacaaagagc tagcctcacg
660
acgcggcaaa taaagagtgc gttgtctgct ataataagatg acaaaaaataa ccocagactcc
720
gtgcacgccg aacactgttt tggattgaa catatattgt gattataatc atcttctaca
780
aatacctag ttacacagct catataacgc aagctagcag acggagtaa accaggggagc
840
cocccacgcc tgcacacagc tgaagccca agacagctgc gacccagcc barccacacagc
900
ggagctgtac atatccaga ggaactgatag tacaasaaac ctttctgtaa aatgaacact
960
gagctgatgct atgcgcaat ccaacagcta aacaaatgtc tlcaocacacct gataaggcttc
1020
gtggaccaag tgtgacagcgg tttgattgagc aatctcagta acaacctctgc ttttcccaca
1080
ttttttccg aggacacagtg tttattctag cgtgcaccca atcggcgcgc acacacaggt
1140
aatgtctca acaaacagcaat ctaacagaaa ggggtttggg tggcaggacac ggaactgtag
1200
catgcctag atattgacca ctaacagggaa tctacacattg aagcacagct cgattttttccc
1260
atcgcgcaac tgtgacagcc aacgtctcac gcacagttga aagatgcgac cgggtctgtag
1320	ttatgcgcc aacaaagaaa cacaactcata accacagctg agaacaacaa gtttgcgcggc
1380
gtggaccaat gttgacagcc aactactcctc atggtctgaa ggcatttgcg cgttaaatttg
1440
acctgtctga ttgattctca tgtgttatca ttatatttg tatttttattttt
1500
tgttacgta tattttcag aagacacggtg atgtgttacaa ataattttgc atttagtaac
1560
taa
1624

<210> SEQ ID NO 17
<211> LENGTH: 897
<212> TYPE: DNA
<213> ORGANISM: respiratory syncytial virus
<400> SEQUENCE: 17
atgctccaaa acaaggaacc acgcaccccc aagcacattag aagggacctg ggaacacctcc
60
aatcttttat tatattttgct atcggattta atataaatgc tgaagcccaaa
120
atctcatttt ctatattcagc aaagatatac tcaacctcag tttataattg acgcacacttca
180
tctatgctc cgacacacca caaagataca cccacacact cgactacata agatgcgaacc
240
agcgagatag aacaacccac cccacactac ccctcaagga atcctcagct tgtgaattagc
300
cctctacatc ctgcttgaa ctactctcata accaccccatc taacactgttc acaaccccca
360
ggagttcact catcctcagct gaccnccca aatcctcgaa cacaacaccg cccaccctcaatc
420
aacaacccca ccaacaccc caacacaccg cccaccctcaatc aacaacaccc
480
aatgattttcg aatcgtttaaag ggaacacctcc atcctcagct gcataagct ccacacacca
540
acetgctttgg attcatcagca aagacatc ccaacaaaa cagcggaaag aacaacctcc
600
agcccacca aacaaacacc cccacagaca cacaacaaaa gcaacatcag cccacacacc
660
aatacagcgg cagcacccca ccaactgagc caccacacta ccaacaaaa aacaacaccc
720
aacaacccca tcaactcata acctcagctgcc aacacaccag gaacacctga aacacactc
780
caaatggaaa cttcctacct ccaacctgctgc aacgcctctc tcaagtctctc
840
aacaacaccc cgcaggtctctc toccacoacc caacacccag ccaggttag
897
What is claimed is:

1. An isolated polynucleotide encoding at least a portion of an infection-attenuated virus of the Pneumoviridae family, the polynucleotide comprising:
   a gene encoding a baculovirus GP64 envelope glycoprotein or variant or fragment thereof; and
   a gene encoding an RSV F protein variant or fragment thereof that comprises at least one amino acid substitution compared to a native RSV F protein, wherein the at least one amino acid substitution stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation; and
   wherein the genome comprises at least one additional modification that renders the virus infection-attenuated.

2. The polynucleotide of claim 1, wherein the at least one amino acid substitution that stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation comprises at least one of the amino acid substitutions S155C, S190F, V207L, and S290C when compared to the native RSV F protein sequence of SEQ ID NO:1.

3. The polynucleotide of claim 1, wherein the RSV F protein variant or fragment thereof comprises the amino acid substitutions S155C, S190F, V207L, and S290C when compared to the native RSV F protein sequence of SEQ ID NO:1.

4. The polynucleotide of claim 1, wherein the virus is further defined as a recombinant respiratory syncytial virus.

5. The polynucleotide of claim 1, wherein the gene encoding the RSV F protein variant or fragment thereof has been codon-optimized.

6. The polynucleotide of claim 1, wherein the gene encoding the RSV F protein variant or fragment thereof comprises at least one of SEQ ID Nos:5-10.

7. The polynucleotide of claim 1, further comprising at least one of:
   a gene encoding an RSV NS1 protein or a variant or fragment thereof;
   a gene encoding an N protein or a variant or fragment thereof;
   a gene encoding a P protein or a variant or fragment thereof;
   a gene encoding an M protein or a variant or fragment thereof;
   a gene encoding an SH protein or a variant or fragment thereof;
   a gene encoding a G protein or a variant or fragment thereof;
   a gene encoding an M-2 protein or a variant or fragment thereof;
   a gene encoding I protein or a variant or fragment thereof; or
   any combination thereof.

8. The polynucleotide of claim 1, wherein:
   (a) a portion of the gene encoding the baculovirus GP64 envelope glycoprotein or variant or fragment thereof encodes an ectodomain of the baculovirus GP64 envelope glycoprotein;
   (b) a portion of the gene encoding the baculovirus GP64 envelope glycoprotein or variant or fragment thereof encodes an ectodomain and a transmembrane domain of the baculovirus GP64 envelope glycoprotein;
   (c) a portion of the gene encoding the baculovirus GP64 envelope glycoprotein or variant or fragment thereof encodes a heterologous cytoplasmic tail; and/or
   (d) at least a portion of the gene encoding the baculovirus GP64 envelope glycoprotein or variant or fragment thereof encodes an amino acid sequence represented by SEQ ID NO:15.
9. A mammalian cell, comprising:
   at least one polynucleotide encoding a baculovirus GP64 envelope glycoprotein or variant thereof;
   and
   at least one polynucleotide encoding at least a portion of a genome of an infection-attenuated virus of the Pneumoviridae family, wherein the genome comprises a gene encoding an RSV F protein variant or fragment thereof that comprises at least one amino acid substitution compared to a native RSV F protein, wherein the at least one amino acid substitution stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation, and wherein the genome comprises at least one additional modification that renders the virus infection-attenuated; and
   wherein the cell produces a recombinant, live, attenuated virus of the Pneumoviridae family.
10. The mammalian cell of claim 9, further defined as a Vero, HEp-2, or 293 cell.
11. The mammalian cell of claim 9, further defined as a Vbac cell.
12. The mammalian cell of claim 9, wherein the at least one polynucleotide encoding a baculovirus GP64 envelope glycoprotein or variant thereof is separate from and not present in the viral genome.
13. The mammalian cell of claim 9, wherein the at least one polynucleotide encoding a baculovirus GP64 envelope glycoprotein or variant or fragment thereof is present in the viral genome.
14. The mammalian cell of claim 9, wherein the at least one amino acid substitution that stabilizes the RSV F protein variant or fragment thereof in a pre-fusion conformation comprises at least one of the amino acid substitutions S155C, S190F, V207L, and S290C when compared to the native RSV F protein sequence of SEQ ID NO:1.
15. The mammalian cell of claim 9, wherein the RSV F protein variant or fragment thereof comprises the amino acid substitutions S155C, S190F, V207L, and S290C when compared to the native RSV F protein sequence of SEQ ID NO:1.
16. The mammalian cell of claim 9, wherein the virus is further defined as a recombinant respiratory syncytial virus.
17. The mammalian cell of claim 9, wherein the gene encoding the RSV F protein variant or fragment thereof has been codon-optimized.
18. The mammalian cell of claim 9, wherein the gene encoding the RSV F protein variant or fragment thereof comprises at least one of SEQ ID NOs:5-10.
19. The mammalian cell of claim 9, wherein the at least one polynucleotide encoding at least a portion of the genome of the infection-attenuated virus of the Pneumoviridae family further comprises at least one of:
   a gene encoding an RSV NS1 protein or a variant or fragment thereof;
   a gene encoding an N protein or a variant or fragment thereof;
   a gene encoding a P protein or a variant or fragment thereof;
   a gene encoding an M protein or a variant or fragment thereof;
   a gene encoding an SH protein or a variant or fragment thereof;
   a gene encoding a G protein or a variant or fragment thereof;
   a gene encoding an M-2 protein or a variant or fragment thereof;
   a gene encoding L protein or a variant or fragment thereof; or
   any combination thereof.
20. The mammalian cell of claim 9, wherein:
   (a) a portion of the polynucleotide encoding the baculovirus GP64 envelope glycoprotein or variant or fragment thereof encodes an ectodomain of the baculovirus GP64 envelope glycoprotein;
   (b) a portion of the polynucleotide encoding the baculovirus GP64 envelope glycoprotein or variant or fragment thereof encodes an ectodomain and a transmembrane domain of the baculovirus GP64 envelope glycoprotein;
   (c) a portion of the polynucleotide encoding the baculovirus GP64 envelope glycoprotein or variant or fragment thereof encodes a heterologous cytoplasmic tail; and/or
   (d) at least a portion of the polynucleotide encoding the baculovirus GP64 envelope glycoprotein or variant or fragment thereof encodes a native amino acid sequence represented by SEQ ID NO:15.
21. A method of producing recombinant, live, infection-attenuated virus of the Pneumoviridae family, the method comprising the steps of:
   culturing the mammalian cell of claim 9 under conditions that allow for production of the recombinant, live, attenuated virus; and
   recovering recombinant, live, infection-attenuated virus.
22. The method of claim 21, further comprising the step of amplifying the attenuated virus in a cell line expressing the baculovirus GP64 envelope glycoprotein or variant or fragment thereof.
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

In the Specification

Column 4, Line 15: Delete “RSV6-preF^{ACT}-AG” and replace with -- RSV6-preF^{ACT,-AG} --

Column 4, Line 36: Delete “fora” and replace with -- for a --

Column 4, Line 46: Delete “RSV-AG-HRP,” and replace with -- RSV-ΔG-HRP, --

Column 10, Line 50: Delete “5190F,” and replace with -- S190F, --

Column 24, Line 10: Delete “RSV6-preF^{ACT,-AG}” and replace with -- RSV6-preF^{ACT,-ΔG} --

Column 24, Line 38: Delete “RSV-AG-HRP,” and replace with -- RSV-ΔG-HRP, --

Signed and Sealed this
Nineteenth Day of March, 2024

Katherine Kelly Vidal
Director of the United States Patent and Trademark Office